

*Journal of Advances in Biology & Biotechnology*

*Volume 27, Issue 10, Page 863-871, 2024; Article no.JABB.120510 ISSN: 2394-1081*

# **Advancements in Rice Biotechnology for Enhanced Abiotic Stress Tolerance**

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#### *Authors' contributions*

*This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.*

#### *Article Information*

DOI:<https://doi.org/10.9734/jabb/2024/v27i101509>

**Open Peer Review History:** This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/120510>

*Review Article*

*Received: 27/07/2024 Accepted: 02/10/2024 Published: 08/10/2024*

#### **ABSTRACT**

Rice (*Oryza sativa* L.) is a vital crop globally, crucial for food security. However, its production faces increasing challenges from abiotic stresses such as drought, cold, heat, salinity, and heavy metals, exacerbated by climate change. This review explores biotechnological approaches aimed at enhancing rice's resilience to these stresses. Key strategies include genetic engineering for introducing stress-tolerant genes, modification of regulatory pathways involved in stress response, and enhancement of physiological adaptations. Advances in biotechnology offer promising avenues for developing rice varieties with improved tolerance to abiotic stress, thereby ensuring sustainable production in diverse agricultural environments.

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*Cite as: Singh, Prabhat Kumar, Shambhoo Prasad, Vandna Kushwaha, Rahul Kumar Maurya, Sumant Pratap Singh, and D. K. Dwivedi. 2024. "Advancements in Rice Biotechnology for Enhanced Abiotic Stress Tolerance". Journal of Advances in Biology & Biotechnology 27 (10):863-71. https://doi.org/10.9734/jabb/2024/v27i101509.*

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*Keywords: Rice; abiotic stress; biotechnology; genetic engineering; stress tolerance.*

#### **1. INTRODUCTION**

"Rice (*Oryza sativa* L.) is a staple cereal crop that feeds half the world's population. Although rice can be grown all over the world, Asian countries account for more than 50% of the total output" [1,2]. "It is a member of the Poaceae family, specifically the genus *Oryza*, which comprises 24 species—22 wild and 2 cultivated" [3]. "Among the cultivated species, *O. glaberrima* and *O. sativa* are well-known, originating from regions across Asia, Europe, the United States, and Africa" [4]. "*O. sativa* is the most widely cultivated rice species due to its adaptability to various regions. It is classified into *japonica*, *indica*, and *javanica* varieties" [4]. *Indica* and *japonica* types are predominantly grown in tropical, subtropical, and temperate regions, while *javanica* is a less common variety adapted to hot and humid conditions.

"Abiotic stressors influence plant development and growth rate at the physiological and biochemical levels, which are essential for increasing crop product efficiency, resulting in losses in yield in the agricultural sector worldwide" [5]. "Different environmental components, such as drought, salinity, heat, cold, and heavy metals, are regarded as important abiotic stresses affecting rice plants. The duration and progression of stress, various stages of plant growth and development, and biotic and abiotic factors may all influence the response to abiotic stresses" [6].

Rice production is significantly affected by diverse abiotic stresses, but these challenges can be mitigated by leveraging genes and regulatory networks from stress-tolerant rice cultivars and other plant species. These genetic resources are instrumental in developing new rice cultivars (see Fig. 1).

Abiotic stresses can activate a diverse array of genes, and intricate transcriptional networks regulate the expression of stress-tolerant genes. Molecular techniques are employed to investigate key genes within these<br>networks. facilitating the development of facilitating the development of transgenic rice varieties that are tolerant to abiotic stresses [7]. As a result, several biotechnology technologies are applied to create rice cultivars that are resistant to abiotic stresses (Table 1).



**Fig. 1. Common abiotic stresses affecting Rice crop**



#### **Table 1. Some transgenes are inserted to improve the abiotic stresses [8]**

### **2. ABIOTIC STRESSES AFFECTING RICE CROP**

#### **2.1 Drought Stress**

"Drought has been affecting agricultural land worldwide for a few years. Many molecular, physiological, and metabolic changes occur in plants due to drought stress that damages their

growth and development" [38]. "During drought stress, plants respond variously and express changes in physiology and morphology. Rice drought resistance is achieved by four procedures that are (a) avoidance: avoiding contact with stress, (b) escape: changing lifecycle, (c) recovery: vegetative growth potency and (d) tolerance: nullifying the impacts of stress. Plants can survive extended periods of drought

and even reproduce in areas with limited water supplies by maintaining physiological activities. These mechanisms include reduced leaf area, leaf rolling, senescence of older leaves, increased root proliferation, dense root system, scavenging reactive oxygen species (ROS), early flowering, osmotic adjustment, stomatal closure that minimizes water loss, changes in the elasticity of cell wall, and maximum uptake of deep water" [39]. Drought stress tolerance in plants can be achieved by accumulating inorganic and organic substances such as proline, potassium ions, glucose, and sucrose. This mechanism, known as osmotic adjustment, keeps the osmotic potential lower inside plant cells than outside, which allows plants to maintain their turgidity and prevent water loss.

#### **2.2 Cold Stress**

"A significant environmental component that has an impact on the development and growth of the rice crop is cold stress. A sudden decrease in temperature may influence the development of chlorophyll during the seedling stage" [40]. "The damage to rice seedlings due to cold stress ultimately decreases the grain yield. So, cold stress is a major limitation that can be overcome by using cold-tolerant rice varieties" [41]. "Because rice crops evolved in tropical regions, it has limited adaptability to chilling stress. Rice cultivation in northern latitudes is made possible by improving rice varieties to make them more cold-tolerant. Chilling tolerance is controlled through many signal transduction pathways and genetic networks" [42]. "In *japonica* rice, chilling tolerance is achieved through interactions between rice G-protein α-subunit 1 (RGA1) and chilling tolerance divergence 1 (COLD1), followed by calcium signaling initiated in the response of the downstream network of stress response that is associated with C repeat binding factor (CBF), a transcription factor" [43]. "However, there is limited information available on the stress response and adaptation. Due to its developmental plasticity, the plant responds to aberrant environmental temperatures by changing its gene expression and adapting to the desirable architecture. Cold stress can disrupt inherent signals in SAMs (shoot apical meristems), and stress tolerance can be increased by regulating the dormancy cycle at the SAM" [44]. "The survival mechanism against cold stress requires the sacrifice of niche forms of root stem cells" [45].

The differentiated cells are well-organized, and this restored development maintains meristematic activity in response to cold temperatures. During cold stress, several particular genes, such as *OsMYB3R-2*, are activated through various transcription factors to maintain mitotic cells and cold tolerance. Survival and growth have been enhanced by maintaining cellular activity and cell function during and after cold stress.

# **2.3 Heat Stress**

"Heat stress is a key limiting factor in agricultural productivity around the world due to global warming. There is a negative correlation between higher temperatures and yields of crops, especially for rice, wheat, barley, and maize" [46]. "Heat stress can severely damage rice plants by decreasing metabolic activity, seed setting, plant growth, and pollen fertility, resulting in reduced rice production" [47]. "Excessive heat can also affect plants' photosynthetic abilities, water use efficiency, seed weight, grain mass, and leaf area. Heat stress can cause damage during both the vegetative and reproductive stages, from sprouting to maturity. However, flowering and booting are the two more essential stages that might result in complete sterility in rice cultivars" [48]. "Heat tolerance refers to plants that are capable of resisting high temperatures while lessening stress and giving enough economic yields. Rice, like other plant species, has genetic variations that help it survive heat stress. Tolerance can be achieved by altering several molecular, morphological, and physiological characteristics in rice cultivars. High temperatures increase the expression of stress-tolerating genes and metabolite reaction, beneficial for plant stress tolerance" [49]. "During heat stress, plants carry out multiple types of responses, including avoidance, survival, and escape. These mechanisms enforce avoidance over the short term and develop resistance for long-term survival. At the cellular level, stress can be controlled by several factors and<br>methods, including transcriptional control, methods, including transcriptional control,<br>antioxidant defense, osmolytes, late antioxidant defense, osmolytes, late embryogenesis abundant (LEA) proteins, and signaling cascade factors. In high temperatures, yield decreases due to early maturity, resulting in comes under the domain of avoidance strategies when it is suffering from heat stress" [48].

# **2.4 Salinity Stress**

"Rice crops are highly vulnerable to salt stress, and approximately one-third of the world's agricultural land is impacted by salinity. The presence of excessive salts in both soil and water detrimentally affects rice production. Rising sodium ion levels in agricultural lands pose a growing threat to global agriculture. This issue causes plants to experience osmotic stress from salt accumulation outside the roots and ionic stress from salt build-up inside the plants" [50]. "The increase in food supply must be equivalent to the rate of increase in population, and this requirement must be satisfied by maximizing the utilization of all available land resources. Therefore, it is also required to enhance the productivity of saline soils. To increase saline soil productivity, different methods such as agronomic adjustments, reclamation, and biological additives are used in combination. Employing genetically improved, salinity-tolerant crop varieties is the best option for achieving sustainable crop production in these regions" [51]. "To develop salt-tolerant crop varieties, it is crucial to evaluate the genetic diversity of crops for salinity tolerance. Molecular mapping techniques have enabled the identification of genomic regions responsible for salt tolerance, making it easier to assess the genetic diversity of various crops and varieties" [52]. "Various molecular mapping techniques can identify the chromosomal regions (QTLs) responsible for salt stress tolerance in rice. Salt stress adversely affects the physiological, morphological, and biochemical characteristics of rice, negatively impacting plant height, shoot dry weight, total tillers, total dry matter, and root dry weight. The physiological attributes affected by salt stress include senescence, uptake of calcium, sodium, and potassium ions, total cation uptake, osmotic potential, transcription efficiency, and relative growth rate" [53]. "Salt stress impacts several biochemical features of rice, including proline content, anthocyanins, peroxidase (POX) activity, calcium content, sodium content, potassium content, chlorophyll content, and hydrogen peroxide content" [50].

#### **2.5 Heavy Metals**

The rhizosphere contains numerous solutes essential for plant growth and development. Plants absorb these solutes through their roots, which then distribute them throughout the entire plant. Successful plant life relies on roots taking up water and other components from the rhizospheric soil. Water uptake, along with soluble elements, drives the developmental plasticity and physiological activity in plant roots. The uptake and distribution of these inorganic materials within plants are fundamental to energy and material fluidity. In plant cells, essential ions

support various physiological and structural functions. However, if these ions are present in non-physiological concentrations, they can become limiting factors. The availability of these ions to plants, disparities in their soil abundance, and their uptake rates affect cellular homeostasis. Plant defense systems and adaptations depend on developmental and physiological changes triggered by ion toxicity, which can also cause permanent damage. The rhizospheric soil also contains heavy metal ions that roots can absorb along with water and nutrients, incorporating them into plant tissues. Toxic metals for plants include zinc, iron, manganese, copper, aluminum, chromium, cadmium, cobalt, lead, arsenic, nickel, and molybdenum [54].

In polluted areas, the concentration of metal ions is excessively high, causing plants to suffer from metal toxicity. Some soils, such as serpentine soils, naturally contain high levels of heavy metals, while mining activities also contribute to elevated heavy metal content in the soil. Environmental pollutants, including high concentrations of heavy metals, are becoming a significant challenge for all organisms—plants, animals, and microbes—worldwide.

#### **3. RICE BIOTECHNOLOGY UNDER CLIMATE CHANGE CONDITIONS**

Abiotic stresses frequently arise from climate change. The impact of these stresses on plant development and yield is evident amidst the changing ecological effects of climate variations [55]. This poses a significant concern for crop production, which has recently increased due to the rapidly rising human population competing for environmental resources [56]. Agriculture, especially rice production, is susceptible to climate change [57]. Abiotic plant stress, encompassing environmental factors like drought, cold, heat, salinity, heavy metals, etc., can ultimately result from severe climatic changes, posing risks to rice crops. Under<br>severe climatic conditions, plants mav severe climatic conditions, plants may experience multiple stresses simultaneously, such as drought and high temperatures, creating unique and unpredictable stress conditions that cannot be anticipated from individual stresses alone [58].

While plants can adapt to changing climatic conditions [59], the simultaneous impact of multiple stressors resulting from frequent climate changes can lead to complete crop failure. A particular environment may be suitable for one plant genotype but can impose various abiotic stresses on another genotype with a different adaptive response [60].

## **4. CONCLUSIONS**

The application of biotechnological tools holds immense promise for addressing the challenges posed by abiotic stresses in rice production. Genetic engineering has shown significant potential in enhancing stress tolerance through the incorporation of stress-responsive genes and the manipulation of regulatory networks. Furthermore, advancements in understanding the physiological mechanisms underlying stress responses offer opportunities for targeted modifications that can improve rice resilience to drought, cold, heat, salinity, and heavy metals stresses. Continued research and development in rice biotechnology are crucial for translating these advancements into practical solutions that ensure sustainable and resilient rice cultivation in the face of evolving environmental conditions.

#### **DISCLAIMER (ARTIFICIAL INTELLIGENCE)**

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

# **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

# **REFERENCES**

- 1. FAO. Statistical pocketbook (Food and Agriculture Organization of the United Nations); 2015.
- 2. Donde R, Gupta MK, Gouda G, Kumar J, Vadde R, Sahoo KK, Behera L. Computational characterization of structural and functional roles of DREB1A, DREB1B and DREB1C in enhancing cold tolerance in rice plant. Amino Acids. 2019;51;839-853.
- 3. Gouda G, Gupta MK, Donde R, Mohapatra T, Vadde R, Behera L. Marker-assisted selection for grain number and yieldrelated traits of rice (*Oryza sativa* L.). Physiol. Mol. Biol. Plants. 2020;26:885- 898.
- 4. Morishima H. Wild plants and domestication. Develo. Crop Sci. 1984;7:3- 30.
- 5. Kazan K. Diverse roles of jasmonates and ethylene in abiotic stress tolerance. Tren. Plant Sci. 2015;20(4):219-229.
- 6. Feller U, Vaseva II. Extreme climatic events: impacts of drought and high temperature on physiological processes in agronomically important plants. Front. Environ. Sci. 2014;2:39.
- 7. Todaka D, Shinozaki K, Yamaguchi-Shinozaki K. Recent advances in the dissection of drought-stress regulatory networks and strategies for development of drought-tolerant transgenic rice plants. Front. Plant Sci. 2015;6:121920.
- 8. Ijaz M, Shahzadi R, Issayeva AU, Bukhari SA. Introductory chapter: recent advances in rice biotechnology for abiotic stress tolerance. Rec. Advan. Rice Res; 2021.
- 9. Jeong MJ, Lee SK, Kim BG, Kwon TR, Cho WS, Park YT, Park SC. A rice (*Oryza sativa* L.) MAP kinase gene, OsMAPK44, is involved in response to abiotic stresses. Plant Cell, Tis. Org. Cul. 2006;85(2):151- 160.
- 10. Agarwal M, Sahi C, Katiyar-Agarwal S, Agarwal S, Young T, Gallie DR, Grover A. Molecular characterization of rice hsp101: Complementation of yeast hsp104 mutation by disaggregation of protein granules and differential expression in indica and japonica rice types. Plant Mol. Biol. 2003;51(4):543-553.
- 11. Oh SJ, Kwon CW, Choi DW, Song SI, Kim JK. Expression of barley HvCBF4 enhances tolerance to abiotic stress in transgenic rice. Plant Biot. J. 2007;5(5): 646-656.
- 12. Wang Q, Guan Y, Wu Y, Chen H, Chen F, Chu C. Overexpression of a rice OsDREB1F gene increases salt, drought, and low temperature tolerance in both Arabidopsis and rice. Plant Mol. Biol. 2008;67(6):589-602.
- 13. Lian HL, Yu X, Ye Q, Ding XS, Kitagawa Y, Kwak SS, Tang ZC. The role of aquaporin RWC3 in drought avoidance in rice. Plant Cell Physiol. 2004;45(4):481-489.
- 14. Shirasawa K, Takabe T, Takabe T, Kishitani S. Accumulation of glycinebetaine in rice plants that overexpress choline monooxygenase from spinach and evaluation of their tolerance to abiotic stress. Annals Bot. 2006;98(3):565-571.
- 15. Zhao FY, Zhang XJ, Li PH, Zhao YX, Zhang H. Co-expression of the Suaeda salsa SsNHX1 and Arabidopsis AVP1 confer greater salt tolerance to transgenic rice than the single SsNHX1. Mol. Breed. 2006;17(4):341-353.
- 16. Feng L, Wang K, Li Y, Tan Y, Kong J, Li H, Zhu Y. Overexpression of SBPase enhances photosynthesis against high temperature stress in transgenic rice plants. Plant Cell Repo. 2007;26(9):1635- 1646.
- 17. Huang CF, Yamaji N, Mitani N, Yano M, Nagamura Y, Ma JF. A bacterial-type ABC transporter is involved in aluminum tolerance in rice. The Plant Cell. 2009; 21(2):655-667.
- 18. Ueno D, Yamaji N, Kono I, Huang CF, Ando T, Yano M, Ma JF. Gene limiting cadmium accumulation in rice. Proc. Natl. Acad. Sci. 2010;107(38):16500-16505.
- 19. Fukao T, Bailey-Serres J. Submergence tolerance conferred by Sub1A is mediated by SLR1 and SLRL1 restriction of gibberellin responses in rice. Proc. Natl. Acad. Sci. 2008;105(43):16814- 16819.
- 20. Huang J, Sun SJ, Xu DQ, Yang X, Bao YM, Wang ZF, Zhang H. Increased tolerance of rice to cold, drought and oxidative stresses mediated by the overexpression of a gene that encodes the zinc finger protein ZFP245. Biochemi. Biophysic. Res. Com. 2009;389(3):556- 561.
- 21. Kumar V, Shriram V, Kavi Kishor PB, Jawali N, Shitole MG. Enhanced proline accumulation and salt stress tolerance of transgenic indica rice by over-expressing P5CSF129A gene. Plant Biot. Repo. 2010; 4:37-48.
- 22. Kim SG, Kim ST, Wang Y, Kim SK, Lee CH, Kim KK, Kang KY. Overexpression of rice isoflavone reductase‐like gene (OsIRL) confers tolerance to reactive oxygen species. Physiol. Planta. 2010; 138(1):1-9.
- 23. Gaxiola RA, Edwards M, Elser JJ. A transgenic approach to enhance phosphorus use efficiency in crops as part of a comprehensive strategy for sustainable agriculture. Chemosphere. 2011;84(6);840-845.
- 24. El-Kereamy A, Bi YM, Ranathunge K, Beatty PH, Good AG, Rothstein SJ. The rice R2R3-MYB transcription factor OsMYB55 is involved in the tolerance to

high temperature and modulates amino acid metabolism. Plos One. 2012; 7(12):e52030.

- 25. Song SY, Chen Y, Chen J, Dai XY, Zhang WH. Physiological mechanisms underlying OsNAC5-dependent tolerance of rice plants to abiotic stress. Planta. 2011; 234(2):331-345.
- 26. Duan J, Cai W. OsLEA3-2, an abiotic stress induced gene of rice plays a key role in salt and drought tolerance. Plos One. 2012;7(9):e45117.
- 27. Gu JF, Qiu M, Yang JC. Enhanced tolerance to drought in transgenic rice plants overexpressing C4 photosynthesis enzymes. Crop J. 2013;1(2):105-114.
- 28. Du H, Wu N, Cui F, You L, Li X, Xiong L. A homolog of ethylene overproducer, O s ETOL 1, differentially modulates drought and submergence tolerance in rice. Plant J. 2014;78(5):834-849.
- 29. Xiong H, Li J, Liu P, Duan J, Zhao Y, Guo X, Li Z. Overexpression of OsMYB48-1, a novel MYB-related transcription factor, enhances drought and salinity tolerance in rice. Plos One. 2014;9(3):e92913.
- 30. Chen H, Liu L, Wang L, Wang S, Cheng X. VrDREB2A, a DREB-binding transcription factor from Vigna radiata, increased drought and high-salt tolerance in transgenic Arabidopsis thaliana. J. Plant Res. 2016;129(2):263-273.
- 31. Cai H, Tian S, Dong H, Guo C. Pleiotropic effects of TaMYB3R1 on plant development and response to osmotic stress in transgenic Arabidopsis. Gene. 2015;558(2):227-234.
- 32. Min HJ, Jung YJ, Kang BG, Kim WT. CaPUB1, a hot pepper U-box E3 ubiquitin ligase, confers enhanced cold stress tolerance and decreased drought stress tolerance in transgenic rice (*Oryza sativa* L.). Mol. Cells. 2016;39(3):250-257.
- 33. Hu T, Zhu S, Tan L, Qi W, He S, Wang G. Overexpression of OsLEA4 enhances drought, high salt and heavy metal stress tolerance in transgenic rice (*Oryza sativa* L.). Environ. Exp. Bot. 2016;123:68-77.
- 34. Shen J, Lv B, Luo L, He J, Mao C, Xi D, Ming F. The NAC-type transcription factor OsNAC2 regulates ABA-dependent genes and abiotic stress tolerance in rice. Sci. Repo. 2017;7(1):40641.
- 35. Park SI, Kim YS, Kim JJ, Mok JE, Kim YH, Park HM, Yoon HS. Improved stress tolerance and productivity in transgenic rice plants constitutively expressing the

*Oryza sativa* glutathione synthetase OsGS under paddy field conditions. J. plant physiol. 2017;215:39-47.

- 36. Li W, Qiang XJ, Han XR, Jiang LL, Zhang SH, Han J, Cheng XG. Ectopic expression of a Thellungiella salsuginea aquaporin gene, TsPIP1; 1, increased the salt tolerance of rice. Int. J. Mol. Sci. 2018;19(8):2229.
- 37. Jiang X, Li S, Ding A, Zhang Z, Hao Q, Wang K, Liu Q. The novel rose MYB transcription factor RhMYB96 enhances salt tolerance in transgenic Arabidopsis. Plant Mol. Biol. Repo. 2018;36(3):406- 417.
- 38. Zu X, Lu Y, Wang Q, Chu P, Miao W, Wang H, La H. A new method for evaluating the drought tolerance of upland rice cultivars. Crop J. 2017;5(6): 488-498
- 39. Saha P, Sade N, Arzani A, Wilhelmi MD MR, Coe KM, Li B, Blumwald E. Effects of abiotic stress on physiological plasticity and water use of *Setaria viridis* (L.). Plant Sci. 2016;251:128-138.
- 40. Kusumi K, Iba K. Establishment of the chloroplast genetic system in rice during early leaf development and at low temperatures. Front. Plant Sci. 2014;5: 101987.
- 41. Zhao J, Zhang S, Dong J, Yang T, Mao X, Liu Q, Liu B. A novel functional gene associated with cold tolerance at the seedling stage in rice. Plant Biot. J. 2017;15(9):1141-1148.
- 42. Zhao C, Lang Z, Zhu JK. Cold responsive gene transcription becomes more complex. Tren. Plant Sci. 2015; 20(8):466-468.
- 43. Zhu JK. Abiotic stress signaling and responses in plants. Cell. 2016;167(2):313- 324.
- 44. Chen L, Zhao Y, Xu S, Zhang Z, Xu Y, Zhang J, Chong K. Os MADS 57 together with Os TB 1 coordinates transcription of its target Os WRKY 94 and D14 to switch its organogenesis to defense for cold adaptation in rice. New Phytol. 2018;218(1):219-231.
- 45. Hong JH, Savina M, Du J, Devendran A, Ramakanth KK, Tian X, Xu J. A sacrificefor-survival mechanism protects root stem cell niche from chilling stress. Cell. 2017; 170(1):102-113.
- 46. Zhang CX, Feng BH, Chen TT, Zhang XF, Tao LX, Fu GF. Sugars, antioxidant enzymes and IAA mediate salicylic acid to

prevent rice spikelet degeneration caused by heat stress. Plant Gro. Regu. 2017; 83(2):313-323.

- 47. Zafar SA, Hameed A, Khan AS, Ashraf M. Heat shock induced morpho-physiological response in indica rice (*Oryza sativa* L.) at early seedling stage. Pak. J. Bot. 2017; 49(2):453-463.
- 48. Zafar SA, Hameed A, Nawaz MA, Wei MA, Noor MA, Hussain M. Mechanisms and molecular approaches for heat tolerance in rice (*Oryza sativa* L.) under climate change scenario. J. Inte. Agri. 2018;17(4):726-738.
- 49. Hasanuzzaman M, Nahar K, Alam MM, Roychowdhury R, Fujita M. Physiological, biochemical, and molecular mechanisms of heat stress tolerance in plants. Int. J. Mol. Sci. 2013;14(5):9643-9684.
- 50. Saeed M. Abiotic stress tolerance in rice (*Oryza sativa* L.): A genomics perspective of salinity tolerance. Rice Crop—Curr. Develop., InTech: London, UK. 2018; 5:181-192.
- 51. Singh R, Singh Y, Xalaxo S, Verulkar S, Yadav N, Singh S, Singh NK. From QTL to variety-harnessing the benefits of QTLs for drought, flood and salt tolerance in mega rice varieties of India through a multiinstitutional network. Plant Sci. 2016; 242:278-287.
- 52. Khan MSK, Saeed M, Iqbal J. Association mapping validates previously identified quantitative trait loci for salt tolerance in rice (*Oryza sativa* L.). Mol. Breed. 2016; 36(12):1-12.
- 53. Negrão S, Schmöckel SM, Tester MJAOB. Evaluating physiological responses of plants to salinity stress. Ann. Bot. 2017; 119(1):1-11.
- 54. Hossain Z, Komatsu S. Contribution of proteomic studies towards understanding plant heavy metal stress response. Front. Plant Sci. 2013;3:41398.
- 55. Bellard C, Bertelsmeier C, Leadley P, Thuiller W, Courchamp F. Impacts of climate change on the future of biodiversity. Ecol. Lett. 2012;15(4):365- 377.
- 56. Wallace JS, Acreman MC, Sullivan CA. The sharing of water between society and ecosystems: from conflict to catchment– based co–management. Philos. Trans. R. Soci. L. Series B: Biol. Sci. 2003; 358(1440):2011-2026.
- 57. Rosenzweig C, Elliott J, Deryng D, Ruane AC, Müller C, Arneth A, Jones JW. Assessing agricultural risks of climate

change in the 21st century in a global gridded crop model intercomparison. Proc. Natl. Acad. Sci. 2014;111(9):3268-3273.

- 58. Suzuki N, Rivero RM, Shulaev V, Blumwald E, Mittler R. Abiotic and biotic stress combinations. New Phytol. 2014; 203(1):32-43.
- 59. Yoshida T, Mogami J, Yamaguchi-Shinozaki K. ABA-dependent and ABA-

independent signaling in response to osmotic stress in plants. Curr. Opin. Plant Biol. 2014;21:133-139.

60. Des Marais DL, Hernandez KM, Juenger TE. Genotype-by-environment interaction and plasticity: exploring genomic responses of plants to the abiotic environment. Annu. Rev. Ecol. Evol. Syst. 2013;44:5-29.

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