



Volume 45, Issue 7, Page 50-58, 2023; Article no.JEAI.98263 ISSN: 2457-0591 (Past name: American Journal of Experimental Agriculture, Past ISSN: 2231-0606)

Spatial Distribution of Spodoptera frugiperda (JE Smith) (Lepidoptera: Noctuidae) on Corn Crops in Durango, Mexico

Claudia Carolina Gurrola-Pérez^a, José Luis Hernández-Mendoza^b, J. Natividad Gurrola Reyes^a and Rebeca Álvarez-Zagoya^{a*}

^a Instituto Politécnico Nacional, Centro Interdisciplinario de Investigación para el Desarrollo Integral Regional, Unidad Durango, Sigma No.119, Fraccionamiento 20 de Noviembre II, Durango, Dgo., México.

^b Instituto Politécnico Nacional, Centro de Biotecnología Genómica, Lab. de Biotecnología Experimental, Blvd. del Maestro s/n esq. Elías Piña, Col. Narciso Mendoza-88710, Reynosa, Tamaulipas, México.

This work was carried out in collaboration among all authors. Author CCGP make experimental work. Authors RAZ and JLHM were advisors and writers. Author RAZ founding and design trails tests. Author JNGR follow the lab works. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JEAI/2023/v45i72131

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/98263

> Received: 18/02/2023 Accepted: 21/04/2023 Published: 04/05/2023

Original Research Article

ABSTRACT

Accurately determining the spatial distribution of an insect is crucial for making effective decisions and efficiently controlling their populations. This study aimed to determine the population distribution of *Spodoptera frugiperda* (J. E. Smith 1797) (Lepidoptera: Noctuidae) larvae in corn

*Corresponding author: E-mail: ralvarezz@ipn.mx, jhernandezm@ipn.mx;

J. Exp. Agric. Int., vol. 45, no. 7, pp. 50-58, 2023

plots near the City of Durango, Mexico. A total of 30 corn-cultivated plots were sampled. In each plot, 5 points were selected (5 of coins method), where 10 consecutive corn plants in these phenological stages V4 to V10 were checked for the presence of larvae and recorded. Statistical analysis was conducted using Scattering Indices $2/\chi^2$ and the Chi-square test (χ^2). The results indicated that infestation was slightly higher during stage V6 than in V4 and V8. The population of *S. frugiperda* had a negative, aggregate, or binomial distribution only in stage V4 (corresponding to small larvae), while in other phenological stages, the distribution was random.

Keywords: Dispersal; crop phenology; maize; statistical analysis.

1. INTRODUCTION

Spodoptera frugiperda is an insect of wide distribution in the American continent and its populations can be detected from Canada to Argentina [1]. This species is reported feeding on more than 200 plant species in the Americas and reported populations have been its in Chikkaballapur, Karnataka in India [2], in Africa where it has spread in 37 countries of the sub-Saharan sector [3-7] and recently in China [8-10]. To better understand Spodoptera frugiperda in India and other countries, it is essential to resume studies on migratory routes, isolation, behavior, alternate hosts, spatial distribution, natural enemies, genetic structure, and habitats for the cold season, as they do not undergo diapause [11-17]. Conducting studies like this can aid in analyzing insect behavior in new distribution areas. For example, in coastal areas of the Pacific, there are overlapping generations until reaching the peak of maximum infestation. However, in the same region with altitudes of 800 to 1200 meters above sea level, a clear generation arrives when the plant is small, followed by overlapping generations that decrease as the plant reaches the reproductive stage [18,19].

The larvae of *S. frugiperda* tend to favor grasses like sorghum, rice, and maize crops. They are typically solitary due to their cannibalistic tendencies [20-22], which impacts their spatial distribution in affected crops [1,15,19,23-26].

Understanding the spatial distribution of this insect is crucial for effective population management decisions [13,27,28]. This is because insect-damaged plants and insect larvae release semiochemicals that parasitoids use to locate their prey [29]. Mathematical models have been employed in studies of spatial distribution to understand the insect's behavior in crops [1,14,30-33]. These models seek to represent the behavior of insect larval populations within a plot and can inform

population management measures, as well as help estimate sample sizes [19,33-35].

The spatial distribution of the noctuid S. frugiperda has been studied, and populations with negative binomial, aggregate, uniform, or random distribution have been reported [19.32.36]. The aggregate distribution is observed in small maize plants in the vegetative phenological stage (V2) or when the larvae hatch before starting their dispersal [18,19]. As the corn plants grow and develop, the larvae tend to separate and remain one per plant until reaching their full growth and pupal stage [15,18,19]. Understanding the biology and ecology of this insect, is of recent relevance due to the infestations it is causing in various locations such as the African continent, China, and Indonesia [3-6,8-10,13,16,17,29,37].

An important aspect to consider is the potential for overlapping generations of *S. frugiperda* in maize plants, which can vary based on factors such as plant phenology, temperature, and the specific maize variety being consumed. To estimate infestations in corn cultivation, a nonlinear regression model can be used based on the distribution of insect larvae within crops [19]. This research was conducted with the goal of improving the management of infestations in maize crops.

2. MATERIALS AND METHODS

The study was conducted in the vicinity of Durango city, on 30 maize cultivation plots located at 24°01'22"N and 104°39'16"W (Fig. 1). Maize cultivation is recommended from May onwards due to the prevailing climatic conditions. The plots were situated at an altitude ranging from 1860 to 1892 meters above sea level. The research team recorded the number of free leaves in 10 continuous plants at 5 different points in each plot. The phenological state of the plants was estimated based on the number of free leaves. The team also checked for the presence of larvae of *S. frugiperda*, noting the number of infested plants and the number of larvae found, following the methods described by Murúa et al. [14] and Hernández-Mendoza et al. [19].

Two methodologies were used to determine the spatial distribution:

a) Variance is obtained by squaring the difference between each observation and the mean, summing up these values, and dividing by n-1. (S2/ x̄. S2 = Variance and x̄ = Population mean. (=((xi - x̄)²)/n-1))

If the values of the dispersion index approach 1, the population's distribution is estimated to be random. If the values are close to zero, a uniform distribution is indicated. Values greater than 1 indicate an aggregate distribution [19,38].

b) Dispersion index obtained with the Chisquare test (χ 2)

If the data obtained are within the values established in the distribution table χ^2 with n-1 degrees of freedom and $\alpha = 0.05$, the distribution is completely random [19]. Excel 2016 was used for the analysis of both indexes. To calculate the indexes, the percentage of infested plants were obtained using the formula proposed by Murúa et al. [14].

3. RESULTS AND DISCUSSION

Upon analyzing the behavior of S. frugiperda infestations detected in this study and their relationship with the phenology of the plant, it becomes evident from Figs.1 and 2, that the infestation rate of this insect in corn plants is relatively low at the beginning and end of crop growth. This low infestation rate may be influenced by the release of the spike, disappearance of the bud, and the insect's own population dynamics [19]. Thus, infestations of S. frugiperda can be detected during all of maize, phenological stages and their populations are influenced by the genetics of the maize plant and the environmental conditions of the site of cultivation [14,19,32].

The sampled sites in this study are located at altitudes greater than 1850 meters above sea level, and the observed infestations were higher

in plants in the phenological stage of V6, when the plants are finishing the vegetative growth stage and until the flag leaf is free, just before the release of the spikes, which is consistent with previously observed behavior in corn grown in sites with altitudes below 500 meters above sea level [18,19]. Similar behavior had also been observed in Argentina [14,15].

In a comprehensive analysis of the sampled sites, considering only the mean population (S2) and the phenological development of *S. frugiperda* larvae (as shown in Table 1), it was found that the highest value was observed in V4, whereas the lowest value was recorded in V10, corresponding to the onset of the reproductive stage of maize, during which the spikes emerge [19].

The data presented above shows a similarity in the variance index values across each phenological stage. This suggests that infestations were high during crop development and decreased as the reproductive stage approached. culminating in the release of the spike. This pattern is consistent with observations made by other authors [14,19].

Fig. 1 illustrates that both $S2/\chi^2$ and larvae (χ^2) exhibit significantly high values in terms of the number of S. frugiperda per sampled plant, and this coincides with the spatial distribution being either of the aggregate or negative binomial type. The determination of population means, and variance alone are not conclusive indicators of the spatial distribution, as demonstrated by this and other studies [33], which is the case for S. frugiperda larvae in maize cultivation under the conditions of this study. Table 1 shows that a sample with V4 has a mean of 0.36, variance of 0.52, an S2/ x2 of 1.45 and an aggregated distribution, whereas another sample with V6 has 0.36, 0.4, and 0.92, respectively, and a random distribution.

An aggregate distribution has also been reported in other insect species during the egg and early developmental stages [32]. This type of distribution is also observed in *S. frugiperda*, where females lay their eggs in aggregate masses with a variable number of eggs. After hatching, the larvae begin to disperse [19].

Vegetative	% Infested	Media	Variance	S2/ <u>x</u>	x2 (Chi	Spatial
State	plants	<u>(x</u>)	(S2)		square)	Distribution
V4	34	0,6	0,9	1,5	11,52	Added
V4	22	0,36	0,52	1,45	16,26	Added
Average	28	0,48	0,71	1,475	13,89	
V6	50	0,64	0,52	0,81	0,62	Random
V6	56	0,7	0,5	0,71	1,21	Random
V6	46	0,56	0,5	0,89	0,4	Random
V6	44	0,48	0,35	0,73	2,75	Random
V6	28	0,28	0,21	0,73	1,19	Random
V6	44	0,52	0,42	0,8	0,52	Random
V6	48	0,6	0,49	0,82	0,45	Random
V6	34	0,36	0,4	0,92	0,07	Random
V6	50	0,58	0,41	0,71	1,82	Random
Average	44.4444	0,52444	0,4222	0,7911	1,00333	
V6-V8	36	0,38	0,28	0,74	2,25	Random
V6-V8	36	0,36	0,24	0,65	2,6	Random
V6-V8	38	0,36	0,23	0,65	3,81	Random
Average	36,6667	0,36667	0,25	0,68	2,88667	
V8	42	0,46	0,34	0,73	2,13	Random
V8	48	0,54	0,38	0,70	2,47	Random
V8	52	0,6	0,47	0,78	2,47	Random
V8	40	0,4	0,25	0,62	5,54	Random
V8	44	0,52	0,42	0,80	0,52	Random
V8	50	0,6	0,46	0,76	0,98	Random
V8	36	0,38	0,3	0,79	2.25	Random
V8	52	0,6	0,41	0,68	2,47	Random
V8	38	0,38	0,24	0,63	5,54	Random
V8	50	0,54	0,34	0,62	5,13	Random
V8	42	0,46	0,34	0,73	2,13	Random
Average	44,9091	0,49818	0,3591	0,7127	2,87545	
V10	42	0,42	0,25	0,59	7,11	Random
V10	26	0,26	0,2	0,76	2,25	Random
V10	22	0,22	0,2	0,89	1,54	Random
V10	44	0,46	0,29	0,64	4,93	Random
Average	33,5	0,34	0,235	0,72	3,9575	

 Table 1. Relationship between the vegetative stages of maize, infested plants and analyses performed to estimate the distribution of *S. frugiperda* larvae



Fig. 1. Result of the application of different methodologies for the estimation of the spatial distribution of larvae of *S. frugiperda* in maize cultivation

The site with the highest infestation was Colonia Hidalgo, and it occurred during vegetative stage V6 (refer to Table 1). Conversely, Hernández-Mendoza et al. [19] reported average infestations of 69% in corn plants during vegetative stage V9 in the state of Colima, which is almost when the plant is fully developed and close to the emergence of the spike. In this case, the infestations caused damage to the corn, resulting in significant losses as the affected crops cannot be sold fresh.

The data presented in Fig. 2 indicates that *S. frugiperda* infestations tend to decrease towards the end of the vegetative development of the crop, consistent with previous findings in maize grown in three agroecological regions of the Mexican Pacific coast (State of Colima), where altitudes are below 1000 meters above sea level [14,19]. This suggests that the insect's behavior is similar at high altitudes as it is at near-sea level [19].

Infested plants suffer more damage from S. frugiperda larvae when the pest occurs in the initial phenological stages than in later vegetative stages. Similarly, Jaramillo et al. [39] mentioned that adults of S. frugiperda prefer early developing maize plants for oviposition. Thus, throughout infestations the vegetative development of corn allow estimating the response or compensation to the loss of foliage caused by insect feeding [18]. This estimation can be made by sampling in any part of the crop, thanks to the random spatial distribution that the insect presents inside it.

Upon analyzing the behavior of *S. frugiperda* infestations detected in this study and its correlation with plant phenology, it becomes apparent that the larvae population reaches its maximum infestation peak when the plant is in full vegetative development, resulting in a parabolic or normal distribution curve (Fig. 2). This behavior has been observed in various eco-geographic conditions and countries where this pest is present, as reported in several studies [1, 14,15,18,19,32,33,39].

Based on the general analysis of the sampled sites in this study, it is evident that the spatial distribution of *S. frugiperda* larvae in the area near Durango city is random, implying that they do not exhibit a defined pattern for infesting plants within a plot. This is important for applications of insecticides or biological control agents, such as the release of parasitoids (Diptera and Hymenoptera), predators, or other agents. Conversely, when the insect exhibits aggregate distribution, management measures must be adjusted, from detection to the application of any form of control.

The spatial distribution of insects can change due to various external factors such as altitudinal and climatic variants, which can vary from year to year [40-46]. However, for the *S. frugiperda* insect, the populations sampled at altitudes above 1800 meters above sea level remain random, just as those at altitudes below 500 meters above sea level have been observed to be random [18,19].



Fig. 2. Estimation of the population behavior of *S. frugiperda* larvae according to the phenological development of corn cultivation in Durango

4. CONCLUSIONS

The spatial distribution of S. frugiperda is estimated to be closely related to the phenology of the maize crop and the present study confirmed that in crops of vegetative development stage V4, that is, in small plants, the distribution is aggregated because they are newly-hatched larvae, while in stages of development V6 to V10 the distribution is random. Thus, control measures with parasitoids or agrochemicals will depend on the age of the plant. The results of the present study may be considered during sampling to determine acceptable levels of infestation.

ACKNOWLEDGEMENTS

To the INSTITUTO POLITÉCNICO NACIONAL, to the SIP-IPN for the financing the project and the work. To CONACYT for the scholarship for Claudia Carolina Gurrola-Pérez. Hernández-Mendoza is SNI and EDI-IPN. Álvarez-Zagoya is supported by EDI-IPN and COFAA-IPN, and Gurrola-Reyes, by COFAA-IPN. We thank the anonymous reviewers for their comments and suggestions.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

 Santos Rios É., Fernandes Martins I.C., Pereira De Noronha M., Alexandre Da Silva J, Gomes Da Silva Filho J., Auguste Badji C. Spatial distribution of *Spodoptera frugiperda* in the wasteland of southern Pernambuco state, Brazil. Cienc. Agr. 2014;57(3):297-304.

DOI: 10.4322/RCA. AO1461.

- ICAR. NBAIR; 2018. Available:http://www.nbair.res.in/recent_ev ents/Pest%20Alert%2030th%20July%2020 18-new1.pdf. [Review date; January 23 2020].
- Goergen G., Kumar P.L., Sankung S.B., Togola A., Tamó M. First report of outbreaks of the fall armyworm *Spodoptera frugiperda* (JE Smith) (Lepidoptera, Noctuidae), a new alien invasive pest in West and Central Africa. Plos One. 2016;11(10):e0165632.

DOI: 10.1371/journal.pone.0165632

- 4. Baudron F, Abdouzaman-allah M, Chaipa I, Newton C, Chinwada P. Understanding the factors influencing fall armyworm (*Spodoptera frugiperda* JE Smith) damage in African smallholder maize fields and quantifying its impact yield. A case study in Eastern Zimbabwe. Crop Prot. 2019; 120:141-50.
- Fotso Kuate AF, Hanna R, Doumtsop Fotio 5. ARP, Abang AF, Nanga SN, Ngatat S. et Spodoptera frugiperda Smith al. (Lepidoptera: Noctuidae) in Cameroon: Case study on its distribution, damage, pesticide use, genetic differentiation and host plants. Plos One. 2019: 14(4):e0215749.

DOI: 10.1371/journal.pone.0215749

- Dahi HF, Salem SA, Gamil WE, Mohamed HO. Heat Requirements for the fall armyworm Spodoptera frugiperda (JE Smith) (Lepidoptera: Noctuidae) as a new invasive pest in Egypt. Egypt Acad. J. Biol. Sci. Entomol. 2020;13(4):73-85.
 DOI: 10.21608/eajbsa.2020.120603
- Mohamed HO, El-Heneidy AH, Dahi HF, Awad AA. First Record of the Fall Armyworm, Spodoptera frugiperda (JE Smith) (Lepidoptera: Noctuidae) on Sorghum Plants, A new invasive pest in Upper Egypt. Egypt Acad. J. Biol. Sci. Entomol. 2022;15(1):15-23.

DOI: 10.21608/eajbsa.2022.214719.

- Wang R, Jiang C., Guo X., Chen D., You C., Zhang Y. et al. Potential distribution of *Spodoptera frugiperda* (J.E. Smith) in China and the major factors influencing distribution. Glob. Ecol. Conserv. 2020;21. DOI: 10.1016/j.gecco.2019.e00865
- Li C, Liao J, Ya Y, Liu J, Li J, Yu G. Analysis of potential distribution of *Spodoptera frugiperda* in western China. J Asia Pac. Entomol. 2022;25(4):101985. DOI: 10.1016/j.aspen.2022.101985
- Jiang YY, Zhang YY, Zhou XY, Hong XY, Chen L. Population genetics reveal multiple independent invasions of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in China. Bull. Entomol. Res. 2022;112(6):796-806.
 DOI: 10.1017/S0007485322000190 [Epub ahead of print]
- López-Edwards M, Hernández-Mendoza JL, Pescador-Rubio A, Molina-Ochoa J, Lezama-Gutiérrez R, Hamm JJ. et al. Biological differences between five

populations of fall armyworm (Lepidoptera: Noctuidae) collected from corn in Mexico. Fla. Entomol. 1999;82(2):254-62. DOI: 10.2307/3496577

- Pashley DP, Johnson SJ, Sparks AN. Genetic population structure of migratory moths: The fall armyworm (Lepidoptera: Noctuidae). Ann. Entomol. Soc. Am. 1985; 78(6):756-62.
 - DOI: 10.1093/aesa/78.6.756
- Westbrook JK, Sparks AN. The role of atmospheric transport in the economic fall armyworm (Lepidoptera: Noctuidae) infestations in the southeastern United States in 1977. Fla. Entomol. 1986;69(3): 492-502.

DOI: 10.2307/3495382.

- 14. Murúa G, Molina-Ochoa J, Coviella C. Population dynamics of the fall armyworm, Spodoptera frugiperda (Lepidoptera: parasitoids Noctuidae) and its in Northwestern Argentina. Fla. Entomol. 2006;89(2):175-82. DOI: 10.1653/0015-4040(2006)89 [175: PDOTFA]2.0.CO;2. TWO. Vol. 2. Available: 2.0.co. DOI: 10.1653/0015-4040(2006)89[175: pdotfa].
- Gabriela Murúa MG, Molina-Ochoa J, Fidalgo P. Natural distribution of parasitoids of larvae of the Fall Army Worm, Spodoptera frugiperda, in Argentina. J. Insect Sci. 2009, 9:20. DOI: 10.1673/031.009.2001
- Wu QL, Jiang YY, Liu J, Hu G, Wu KM. Trajectory modeling revealed a southwestnortheast migration corridor for fall armyworm *Spodoptera frugiperda* (Lepidoptera: Noctuidae) emerging from the North China Plain. Insect Sci. 2021;28(3):649-61.

DOI: 10.1111/1744-7917.12852

17. Afandhi A, Fernando I, Widjayanti T, Maulidi AK, Radifan HI, Setiawan Y. Impact of the fall armyworm, Spodoptera frugiperda (J.E. Smith) (Lepidoptera: Noctuidae), invasion on maize and the native Spodoptera litura (Fabricius) in east Java, Indonesia, and evaluation of the virulence of some indigenous entomopathogenic fungus isolates for controlling the pest. Egypt J. Biol. Pest Control. 2022;32(1):48.

DOI: 10.1186/s41938-022-00541-7

 Hernández-Mendoza JL. Ecopathologie et degats de Spodoptera frugiperda (J.E. Smith) sur la culture du maize in Mexico [Ph.D. thesis]. State of Colima - Possibilitees de lutte a l'aide de la bacterie entomopathongene *Bacillus thuringiensis*. Université des Sciences, du Languedoc T. Francia. 1989;142.

 Hernández-Mendoza JL., López-Barbosa EC, Garza-González E, Mayek-Pérez N. Spatial distribution of Spodoptera frugiperda (Lepidoptera: Noctuidae) in maize landraces grown in Colima, Mexico. Int. J. Trop. Insect Sci. 2008; 28(3):126-9.

DOI: 10.1017/S1742758408096112

20. Silva CSBD, Parra JRP. New method for rearing *Spodoptera frugiperda* in laboratory shows that larval cannibalism is not obligatory. Rev. Bras. Entomol. 2013;57(3):347-9.

DOI: 10.1590/S0085-56262013005000029 21. Andow DA, Farias JR, Horikoshi RJ,

- Bernardi D, Nascimento ARB, Omoto C. Dynamics of cannibalism in equal-aged cohorts of *Spodoptera frugiperda*. Ecol. Entomol. 2015;40(3):229-36. DOI: 10.1111/een.12178.
- 22. Andow DA, Farias JR, Horikoshi RJ, Bernardi D, Nascimento ARB, Omoto CO. Dynamics of cannibalism in equal-aged cohorts of *Spodoptera frugiperda*. Ecol. Entomol. 2015;40(3):229-36. DOI: 10.1111/een.12178.
- Barfield CS, Ashley TR. Effects of corn phenology and temperature on the life cycle of the fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae). Fla. Entomol. 1987;70(1):110-6. DOI: 10.2307/3495097.
- Simmons AM, Marti OG. Mating by the fall armyworm (Lepidoptera: Noctuidae): Frequency, duration, and effect of temperature. Environ. Entomol. 1992; 21(2): 371-5. DOI: 10.1093/ee/21.2.371

 Casmuz T, Juarez ML, Socías MG, Murúa MG, Prieto S, Medina S, et al. Review of the hosts of fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae). Rev. Soc. Entomológica Argent. 2010; 69(3-4):209-31.

- 26. Bohnenblust E, Tooker J. Fall armyworm as a pest of field corn; 2012.
 Available: https://extension.psu.edu/fall-armyworm-as-a-pest-of-field-corn.
 [Review date: January 23, 2020]
- 27. Giles KL., Royer TA, Elliott NC, Kindler SD. Development and validation of a binomial sequential sampling plan for the

greenbug (Homoptera: Aphididae) infesting winter wheat in the southern plains. J. Econ. Entomol. 2000;93(5):1522-30.

DOI: 10.1603/0022-0493-93.5.1522

 Yanqui Diaz F, Camacho JA, Ustua HC, Caballero Ramírez SC, Carrasco BS, Bocanegra DEC, et al. Umbral de tratamiento del gusano cogollero (*Spodoptera frugiperda*) en el cultivo de maíz amiláceo (*Zea mays* L. ssp. amiláceo). Manglar. 2022;19(3): 291-7.

DOI: 10.17268/manglar.2022.037

Mohammed K, Agarwal M, LI B, Newman J., Liu T., Ren Y. Evaluation of d-limonene and β-ocimene as Attractants of *Aphytis melinus* (Hymenoptera: Aphelinidae), a Parasitoid of *Aonidiella aurantii* (Hemiptera: Diaspididae) on *Citrus* spp. Insects. 2020; 11(1):44.

DOI: 10.3390/insects11010044

 Terry I, Bradley JR, Van Duyn JW. Heliothis zea (Lepidoptera: Noctuidae) eggs in soybeans: within-field distribution and precision level sequential count plans. Environ. Entomol. 1989;18(6): 908-16.

DOI: 10.1093/ee/18.6.908

- Álvarez J, Martinez O. Sequential sampling plan for larvae of fall armyworm (Lepidoptera: Noctuidae), in maize. Colomb. Agron. 1990;7(1):32-6.
- 32. Farias-Rivera L.A., Hernández-Mendoza J.L., Molina-Ochoa J., Pescador-Rubio A. Effect of leaf extracts of teosinte, *Zea diploperennis* L., and a Mexican maize variety, criollo "Uruapeño", on the growth and survival of the fall armyworm (Lepidoptera: Noctuidae). Fla. Entomol. 2003, 86(3):239-43.

DOI: 10.1653/0015-4040(2003)086[0239:EOLEOT]2.0.CO;2

- Crespo-Herrera LA, Vera-Graziano J, Bravo-Mojica H, López-Collado J, Reyna-Robles R, Peña-Lomelí A, et al. Spatial distribution of *Bactericera cockerelli* (Sulc) (Hemiptera: Triozidae) on green Tomato (*Physalis ixocarpa* (Brot). Agrociencia. 2012;46(3):289-98.
- 34. Southwood T.R.E. Ecological methods with particular reference to the study of insect populations. London: Methuen and Co., Ltd.; 1966.

- Myers JH. Selecting a measure of dispersion. Environ. Entomol. 1978;7(5): 619-21.
 DOI: 10.1093/ee/7.5.619
- 36. Clavijo S. Spatial distribution of fall armyworm *Spodoptera frugiperda* (Smith) (Lepidoptera: Noctuidae). J. Fac. Agron. (Luz, Venezuela). 1978;26:101-6.
- Nagoshi RN, Fleischer S, Meagher RL, 37. Texas is the overwintering source of fall armvworm in central Pennsvlvania: implications for migration into the northeastern United States. Environ. Entomol. 2009;38(6):1546-54. DOI: 10.1603/022.038.0605
- Vivas LE, Notz A. Spatial distribution in populations of *Oebalus insularis* Stal (Hemiptera: Pentatomidae) in rice cultivation in Calabozo, Guárico state, Venezuela. Rev. Cient. UDO Agric. 2011; 11(1):109-25.
- Jaramillo T, Jaramillo O, Bustillo T, Gómez H. Effect of fall armyworm *Spodoptera frugiperda* (J.E. Smith) on maize yield. Rev. Fac. Nac. Agron. 1989;42(1):25-33.
- 40. Redolfi I, Ruano F, Tinaut T, Pascual F, Campos M. Spatial distribution and temporal permanence of anthills in the olive tree agrosystem in Granada, Spain. J. Appl. Ecol. 2005;4(1, 2):71-76.
- Alonso-Hernández N, Sánchez-García JT, Figueroa De La Rosa JI, López-Martínez V, Martínez-Martínez L, Pérez-Pacheco R, et al. Spatial distribution of braconids (Hymenoptera) reported in the state of Oaxaca. Acta Zoológica Mex. 2014; 30(3):564-94.
- Amell-Caez YNA, De Castro-Arrazola I, García H, Monroy G, Noriega JA. Spatial diversity of dung beetle assemblages (Coleoptera: Scarabaeidae: Scarabaeinae) in five ecoregions from Sucre, Colombian Caribbean coast. Rev. Colomb. Entomol. 2019;45(2):e7963.

DOI: 10.25100/socolen.v45i2.7963

- 43. Castillo A. Corn. In: Agricultural Technical Agenda Durango. SENASICA-SAGARPA I. 2015;176: 91-101. Available:https://issuu.com/senasica/docs/ 10_durango_2015_sin
- Hailu G, Niassy S, Bässler T, Ochatum N, Studer C, Salifu D, et al. Could fall armyworm, Spodoptera frugiperda (J.E. Smith) invasion in Africa contribute to the displacement of cereal stemborers in

maize and sorghum cropping systems?. Int. J. Trop. Insect Sci. 2021;41(2): 1753-62.

DOI: 10.1007/s42690-020-00381-8.

45. Ritchie SW, Hanway JJ, Benson GO, Herman JC. How a corn plant develops. SPECIAL REPORT Num.48. Iowa State University of Science and Technology Coop. Ext. Serv. Ames, IA; 1992.

 Westbrook JK, Nagoshi RN, Meagher RL, Fleischer SJ, Jairam S. Modeling seasonal migration of fall armyworm moths. Int. J. Biometeorol. 2016;60(2):255-67. DOI: 10.1007/s00484-015-1022-x

© 2023 Gurrola-Pérez et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

> Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/98263