

British Journal of Pharmaceutical Research 3(1): 46-53, 2013

SCIENCEDOMAIN *international www.sciencedomain.org*

Rhoifolin; A Potent Antiproliferative Effect on Cancer Cell Lines

Omayma A. Eldahshan1*

¹Pharmacognosy Department, Faculty of Pharmacy, Ain Shams University, Cairo, Egypt.

Author's contribution

The author designed the study, performed the data analysis and wrote the manuscript. He read and approved the final manuscript.

Research Article

Received 22nd July 2012 Accepted 5 th November 2012 Published 19 th December 2012

ABSTRACT

Aims: To investigate the cytotoxic activity of rhoifolin against different cancer cell lines. **Study Design:** Isolation, identification and cytotoxic activity evaluation.

Place and Duration of Study: Faculty of Pharmacy, Ain Shams University and Al-Azhar University, between October, 2010 and January, 2011.

Methodology: Rhoifolin, Apigenin 7-O-β neohesperidoside was isolated in a copious amount from the leaves of *Chorisia crispiflora* (Bombaceae). Its identity was unambiguously confirmed via different spectroscopic methods (UV, ¹HNMR, ¹³CNMR and HMBC) and viability assay test was used to evaluate its cytotoxic activity.

Results: It exhibited potent anticancer activities, nearly similar to that of vinblastine, when evaluated against human epidermoid larynex (Hep 2) and human cervical (HeLa) carcinoma cell lines. Promising activities were also obtained against hepatocellular (Hep G2), colon (HCT-116) and fetal human lung fibroblast (MRC-5) carcinoma cell lines. A unique effect of rhoifolin was in having no cytotoxic activity against healthy normal cells (Vero cells) which indicates a high selectivity of this compound.

Conclusion: The findings of this study showed that rhoifolin could be used as an ideal anticancer agent. It discriminates between cancerous and non cancerous cell as it kills only the former one. So the side effects which may appear during chemotherapy could be overcome.

__

^{}Corresponding author: Email: omiahm@hotmail.com;*

Keywords: Roifolin; Chorisia crispiflora; Bombaceae; spectroscopic methods; cytotoxic activity.

1. INTRODUCTION

The introduction of active agents derived from nature into the cancer armamentarium has changed the natural history of many types of human cancer. Throughout medical history, novel plant-derived compounds were of great significance to cancer therapy. As examples of these compounds are vinblastine and vincristine; *Catharanthus roseus* family Apocynaceae [1]. Currently, one of the major cancer treatments is chemotherapy. Most of the chemotherapeutic drugs such as vincristine, paclitaxel, and etoposide (ET) cannot discriminate between cancer and non-cancer cells. Many normal cells are also killed during the process of chemotherapy. This nonspecific cytotoxicity damages the patient's immune system and generates many side effects such as neutropenia, vomiting, hair loss, peripheral neurotoxicity, etc. [2, 3].

Rhoifolin is apigenin 7-O-β neohesperidoside. It was reported that rhoifolin has lots of pharmacological actions. It exerts its anti-diabetic effect through enhanced adiponectin secretion, phosphorylation of insulin receptor-*β*, and GLUT4 translocation [4]. It has an antiinflammatory action via multi-level regulation of inflammatory mediators [5]. Rhoifolin produced no change in hypoxic pulmonary vasoconstriction, but decreased cardiac output and aortic pressure [6].

Apigenin nucleus is a cancer chemopreventive agent. It inhibits cell proliferation in cancer cell types [7]. Because of its potential antioxidant, anti-inflammatory, and anti-tumor properties, apigenin is considered as a candidate cancer chemopreventive agent [8-11].

Apigenin inhibited the growth through an apoptotic pathway in human cervical carcinoma HeLa cells [12]. Furthermore, apigenin inhibited A549 lung cancer cell proliferation and vascular endothelial growth factor (VEGF) transcriptional activation in a dose-dependent manner [13]. However, Kawaii [14] reported the weak effect of rhoifolin against melanin pigment producing mouse melanoma, human T-cell leukemia, and human lung and lymph node metastatic carcinoma cell lines.

So far, nothing has been documented about the cytotoxic effect of rhoifolin against human epidermoid larynex (Hep 2), human cervical (HeLa), hepatocellular (Hep G2), colon (HCT- 116) and fetal human lung fibroblast (MRC-5) carcinoma cell lines, so the antineoplastic activity of this compound was investigated against these types of cancer cell lines. We reported here for the first time the high potent and selective antitumor activity of rhoifolin against several types of cancer cell lines to develop a preliminary building block for the construction of a new anticancer drug.

2. MATERIALS AND METHODS

2.1 Plant Material

Chorisia leaves were collected from Zoo Garden in Giza, Egypt, 2010 and were authenticated by Prof. Dr Abdel Salam El Noyehy, Prof. of Taxonomy, Faculty of Science, Ain Shams University, Cairo, Egypt. Voucher specimen was deposited in the herbarium of Pharmacognosy Department (voucher specimen number; CCB-73), Faculty of Pharmacy,

Ain Shams University, Cairo, Egypt. The leaves were dried in shade and milled to a fine powder.

2.2 Extraction and Isolation

Powder of air dried leaves of *Chorisia crispiflora* (1 kg) was extracted with 70 % ethanol at room temperature. The extract was entirely dried and dissolved in a small amount of water and partioned with *n*-hexane, ethyl acetate and butanol successively. The aqueous water residue was totally dried and extracted with methanol at 40ºC. The methanolic extract upon concentration yielded yellow crystals of rhoifolin (8.3 g). Purification to the crystals was achieved by crystallization.

2.3 Instruments and Materials for Phytochemical Investigation

Chromatographically pure materials 1 mg each were dissolved in analytically pure methanol next subjected to UV spectroscopic investigation in 4 ml capacity quartz cells 1 cm thick using a Carl Zeiss spectrophotometer PMQ II. AlCl₃, AlCl₃/HCl, fused NaOAc / H₃BO₃ and NaOMe reagents were separately added to the methanolic solution of investigated material and UV measurements were later carried out.

The NMR spectra were recorded on a Varian Mercury VX-500 NMR spectrometer. ¹Hspectra ran at 300 MHz and 13 C- spectra were run at 75.46 MHz in deutrated dimethylsulphoxide (DMSO-*d6*).

Rhoifolin: Apigenin 7-O-β neohesperidoside, $C_{27}H_{30}O_{14}$, yellow needles, m.p. 250-265°C. IR νmax (KBr): 3388 (OH), 1657 (α, β-unsat. CO), 1605, 1497, and 1488 (arom. C=C), 1249, 1178, 1074 (glycosidic C–O) cm-1 .

UV λmax (log ɛ) (MeOH): 266 (4.20), 336 (4.30) nm; (MeONa): 267 (4.20), 387 (4.40) nm; (NaOAc): 257 (4.20), 266 (4.20), 391 (4.40) nm, (NaOAc + H₃BO₃): 268 (4.20), 340 (4.30) nm; (AlCl₃): 275 (4.20), 299 (4.10), 350 (4.20), 385 (4.20) nm, (AlCl₃ + HCl): 276 (4.20), 298 (4.10), 342 (4.20), 382 (4.10) nm.

¹H-NMR, DMSO-*d⁶* δ ppm: 7.91(2H, d, *J*=8.8 Hz, H-2`,6`), 6.92 (2H, d, *J*= 8.8 Hz, H-3`,5`), 6.84 (1H, d, *J*= 2.0 Hz, H-8), 6.80 (1H, s, H-3), 6.33 (1H, d, *J*= 2.0 Hz , H-6), 5.08 (1H, singlet like, H-1```), 5.20 (1H, d, *J*= 7.3 Hz, H-1``), 1.16 (3H, d, *J*=6.3Hz,CH3-6```).

¹³C-NMR, DMSO-*d⁶* δ ppm: 182.1-C4, 164.4-C2, 162.6-C7, 161.7-C4`, 161.1-C5, 157.1-C9, 128.7-C2`,6`, 120.9-C1`, 116.2C-3`,5`, 105.5-C10, 103.2-C3, 99.4-C6, 94.6-C8, Sugar proton: 100.5-C1``, 98.2-C1```, 77.6-C2``, 77.4-C3``, 76.8-C5``, 72.3-C4```, 71.0-C2```, 70.8- C3```, 71.1-C4``, 68.8-C5```, 60.9-C-6``, 18.5-C-CH₃.

2.4 Mammalian Cell Lines

Vero cells (Normal kidney cells).

Carcinoma cell lines: Hep2 (human epidermoid larynex carcinoma cells), HeLa cells (human cervical carcinoma cells), Hep G2 (human hepatocellular carcinoma), HCT-116 (human colon carcinoma cells) and MRC-5 (fetal human lung fibroblast cells).

All cell lines of a well-differentiated carcinoma were obtained from the American Type Culture Collection (ATCC).

2.5 Chemical Used

Dimethyl sulfoxide (DMSO), crystal violet and trypan blue dye (Sigma, St.Louis, Mo., USA).

DMEM, RPMI-1640, FBS, HEPES buffer solution, L-glutamine, gentamycin and 0.25 % Trypsin-EDTA (Bio Whittaker ®Lonza, Belgium). Crystal violet stain (1%).

2.6 Cytotoxicity Evaluation Using Viability Assay

Cell toxicity was monitored by determining the effect of the test sample on cell viability through the viability test [15, 16].

3. RESULTS AND DISCUSSION

3.1 Identification of the Compound

Pure material of rhoifolin was obtained as an amorphous light yellow powder, which appeared as a dark purple spot on Paper chromatography (PC) and turned yellow upon exposure to ammonia vapors, under short UV light (254 nm). Confirmation of the compound was achieved through UV shift reagents, 1 HNMR, 13 CNMR and HMBC correlation (Fig. 1).

3.2 Cytotoxic Activity of the Compound

The tested compound showed marked toxic effects to the cancerous cell lines. It exerted cytotoxic activity to Hep 2 and HeLa cell lines at IC_{50} 5.90 and 6.2 μ g/mL respectively (Fig. 2 B and C). HepG2 is affected but to a lesser extent by the compound at IC_{50} 22.6 µg/mL (Fig. 2-D). The least potent activities were to HCT-116 and MRC-5 at IC_{50} 34.8 and 44.6 μ g/mL respectively (Fig. 2 E and F).

Historically natural products have been an important source of antineoplastic drugs. Sixty percent of currently used antitumor agents are of natural origin, derived from plants, marine organisms and is a useful tool for the discovery of new potential anticancer agents from natural products. One of the important criteria for a therapeutic drug for cancer is to have minimum or no side effects on normal body cells of patients undergoing chemotherapy. This invariably implies that the drug should not only have high potent activity at lower concentrations but also should exhibit a high degree of selectivity.

Thus, development of novel selective drugs is an important and challenging task, and understanding the biological differences between normal and cancer cells is essential for achieving this goal.

The present *in vitro* study showed the ability of rhoifolin to exhibit a high degree cytotoxic activity to cancerous cells with great selectivity, where, as it is clear, that the compound has no cytotoxic activity against mammalian normal cells (Table 1, Fig. 2A).

British Journal of Pharmaceutical Research, 3(1): 46-53, 2013

Fig. 1. HMBC correlations of apigenin 7-*O***-β neohesperidoside (Rhoifolin)**

Table 1 showed that the compound of interest exhibited high cytotoxic activity to laryngeal cancer cells, at a very low IC50= 5.9 µg/mL followed by cervical at IC50= 6.2 µg/mL, both are nearly similar to that of vinblastine. Hepatic cell line was also affected but at a lesser extent by the toxicity of rhoifolin at IC50 22.6 µg/mL. The colon and the fetal human lung fibroblast cell lines are affected at IC50 34.8 and 44.6 µg/mL respectively.

The selectivity index (SI) was defined as the ratio of the IC50 obtained from the experiment on normal cells vs. cancer cells. High selectivity was achieved when the SI was ≥ 3 [17]. As the Selective index (SI) demonstrates the differential activity of a pure compound, the greater the SI value is, the more selective it is. An SI value less than 2 indicates general toxicity of the pure compound [18].

Based on this, the SI data shown in Table 2 indicates that rhoifolin exhibits a very high degree of cytotoxic selectivity at SI greater than 8.47 for laryngeal cell lines, followed by 8.06 in cervical and 2.21 in hepatic carcinoma cell lines. The other two carcinoma cell lines; colon and fetal human lung fibroblast are of little SI.

Table 1. IC50 of rhoifolin and vinblastine on the carcinoma cell lines

Fig. 2. Cytotoxic activity of rhoifolin on different human cell lines (the concentration in µg/mL): (A) Mammalian vero cell line (normal cell), (B) Hep2 (human epidermoid larynex carcinoma cells), (C) HeLa cells (human cervical carcinoma cells), (D) Hep G2 (human hepatocellular carcinoma), (E) HCT-116 (human colon carcinoma cells) and (F) MRC-5 (fetal human lung fibroblast cells)

4. CONCLUSION

Interestingly, this present study, showed the following advantages of rhoifolin:

1. Potent cytotoxic effect nearly similar to that of vinblastine which may become a good therapeutic strategy to its use as an antagonist for treatment of this dreaded disease, especially laryngeal, cervical and hepatic cancer.

2. It is considered as an ideal antitumor agent to specific cancerous cells where it is toxic to malignant with no toxicity to normal cells so it will be a good building unit for a new antitumor drug without side effects.

However, currently there are limited numbers of such agents available for clinical use. The mechanisms behind its respective anticancer effect are now under investigation to pave a way to a discovery of a new cancer therapeutic agent.

ACKNOWLEDGEMENTS

The author is grateful for The Regional Center for Mycology and Biotechnology, Al-Azhar University, Cairo, Egypt for hosting the cytotoxic activities.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

- 1. Kalidass C, Mohan VR, Daniel A. Effect of auxin and cytokinin on vincristine production by callus cultures of *Catharanthus roseus* L. (apocynaceae). Trop. Subtrop. Agroecosystems. 2010;12:283-288.
- 2. Perry MC, Moertel CG, Schutt AJ, Reitemeier RJ, Hahn RG. Phase II studies of dianhydrogalactitol and VP-16-213 in colorectal cancer. Cancer Treat Rep. 1976;60: 1247–50.
- 3. Einzig AI, Hochster H, Wiernik PH, Trump DL, Dutcher JP, Garowski E. A phase II study of taxol in patients with malignant melanoma. Invest New Drugs. 1991;9:59–64.
- 4. Rao YK, Lee M, Chen K, Lee Y, Wu W, Tzeng W. Mimetic Action of Rhoifolin and Cosmosiin Isolated from *Citrus grandis* (L.) Osbeck Leaves: Enhanced Adiponectin Secretion and Insulin Receptor Phosphorylation in 3T3-L1 Cells. Evidence-Based Complementary and Alternative Medicine. 2011:9.
- 5. Eldahshan OA, Azab SS. Anti-inflammatory Effect of Apigenin-7- neohesperidoside (Rhoifolin) in Carrageenin- Induced Rat Oedema Model. Journal of Applied Pharmaceutical Science. 2012;02(08):74-79.
- 6. Occhiuto F, Limardi F. Comparative effects of the flavonoids luteolin, apiin and rhoifolin on experimental pulmonary hypertension in the dog. Phytotherapy Research. 1994;8(3): 153–56.
- 7. Sarkar FH, Li Y. Cell signaling pathways altered by natural chemopreventive agents. Mutat. Res. 2004;555:53–64.
- 8. Birt DF, Pelling JC, Nair S, Lepley D (1996). Diet intervention for modifying cancer risk. Prog. Clin. Biol. Res. 1996;395:223–34.
- 9. Birt DF, Mitchell D, Gold B, Pour P, Pinch HC. Inhibition of ultraviolet light induced skin carcinogenesis in SKH-1 mice by apigenin, a plant flavonoid. Anticancer Res.1997;17:85–91.
- 10. Lepley DM, Li B, Birt DF, Pelling JC. The chemopreventive flavonoid apigenin induces G2/M arrest in keratinocytes. Carcinogenesis. 1996;17:2367–75.
- 11. Ross JA, Kasum CM. Dietary flavonoids: bioavailability, metabolic effects, and safety. Annu. Rev. Nutr. 2002;22:19–34.
- 12. Zheng PW, Chiang LC, Lin CC. Apigenin induced apoptosis through p53-dependent pathway in human cervical carcinoma cells. Life Sci. 2005;76:1367–79.
- 13. Liu LZ, Fang J, Zhou Q, Hu X, Shi X, Jiang BH. Apigenin inhibits expression of vascular endothelial growth factor and angiogenesis in human lung cancer cells:implication of chemoprevention of lung cancer. Mol Pharmacol. 2005;68(3):635- 43.
- 14. Kawaii S, Tomono Y, Katase E, Ogawa K, Yano M. Antiproliferative activity of flavonoids on several cancer cell lines. Biosci Biotechnol Biochem. 1999;63(5):896-99.
- 15. Vijaya P, Raghu C, Ashok G, Dhanaraj SA, Suresh B. Antiviral activity of medicinal plants of Nilgiris. Indian J Med. Res. 2004;120:24-29.
- 16. Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. J Immunol. Methods. 1993;65:55-63.
- 17. Prayong P, Barusrux S, Weerapreeyakul N. Cytotoxic activity screening of some indigenous Thai plants. Fitoterapia. 2008;79:598-601.
- 18. Koch A, Tamez P, Pezzuto J, Soejarto D. Evaluation of plants used for antimalarial treatment by the Massai of Kenya. J Ethnopharmacol. 2005;101:95-99.

___ *© 2013 Eldahshan; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.*

Peer-review history: The peer review history for this paper can be accessed here: http://www.sciencedomain.org/review-history.php?iid=172&id=14&aid=802