

Asian Journal of Research and Reports in Neurology

Volume 7, Issue 1, Page 154-164, 2024; Article no.AJORRIN.124031

# Ethanol Extract of Stachytarpheta jamaicensis Reduces Lifespan Shortening, Oxidative Stress, and **Neurotoxicity in Transgenic Drosophila** Parkinson's Disease Model

Theresa E. Isamoh <sup>a\*</sup>. Francis Peter Ikumosam <sup>a</sup>. John Afeez Olanrewaju<sup>b</sup>, Emmanuel I. Odom<sup>a</sup>, Eru, Eru Mba <sup>a</sup>, Nsikak Michael Umoh <sup>a</sup>, Sadeyeng Ernest Anani<sup>a</sup> and Ibharale Felicity Onose<sup>c</sup>

<sup>a</sup> Department of Anatomical Sciences, Faculty of Basic Medical Sciences, University of Calabar, Nigeria.

<sup>b</sup> Department of Anatomy, Faculty of Basic Medical Sciences, Babcock University, Nigeria. <sup>c</sup> Ambrose Alli University, Ekpoma, Edo State, Nigeria.

#### Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

#### Article Information

**Open Peer Review History:** This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/124031

> Received: 26/07/2024 Accepted: 28/09/2024 Published: 03/10/2024

**Original Research Article** 

\*Corresponding author: E-mail: theresaisamoh@yahoo.com;

Cite as: Isamoh, Theresa E., Francis Peter Ikumosam, John Afeez Olanrewaju, Emmanuel I. Odom, Eru, Eru Mba, Nsikak Michael Umoh, Sadeyeng Ernest Anani, and Ibharale Felicity Onose. 2024. "Ethanol Extract of Stachytarpheta Jamaicensis Reduces Lifespan Shortening, Oxidative Stress, and Neurotoxicity in Transgenic Drosophila Parkinson's Disease Model". Asian Journal of Research and Reports in Neurology 7 (1):154-64.

https://www.journalajorrin.com/index.php/AJORRIN/article/view/112.

# ABSTRACT

**Background:** Parkinson's disease (PD) is a neurodegenerative disorder characterized by the progressive death of dopaminergic neurons. Stachytarpheta jamaicensis, a medicinal plant, has been shown to possess neuroprotective effects. The medicinal plant has been traditionally used to treat various ailments, including neurological disorders. This study evaluated the effect of S. jamaicensis leaf extract on a transgenic Drosophila model of PD.

**Methods:** Five groups of Drosophila melanogaster were treated with S. jamaicensis extract: wild control, wild + treatment, transgenic control, transgenic + low dose (50mg/kg), and transgenic + high dose (250mg/kg). Flies were cultured and treated for five days before behavioral, lifespan, and biochemical testing.

**Results:** S. jamaicensis extract increased lifespan and locomotor activity in flies. Biochemical assays revealed reduced acetylcholine esterase activity, oxidative stress markers (malondialdehyde and total oxidized protein), and increased catalase enzyme activity and total thiol levels, p< 0.05; p< 0.01; p< 0.005.

**Conclusion:** S. jamaicensis leaf extract exhibited neuroprotective and antioxidant effects in the Drosophila melanogaster model of PD, potentially through modulation of oxidative stress pathways. These findings suggest that S. jamaicensis extract may be a potential therapeutic agent for PD treatment.

Keywords: Parkinson's disease; Stachytarpheta jamaicensis; Neuroprotection; antioxidant; oxidative stress; Drosophila melanogaster; Transgenic model; Neurodegenerative disorder.

### 1. INTRODUCTION

Parkinson's disease (PD) is a progressive neurodegenerative disorder characterized by the selective loss of dopaminergic neurons in the substantia nigra pars compacta (SNpc) that project to the striatum [1]. The clinical diagnosis of PD is based on dopamine deficiency, and alterations in neurotransmitter levels are also known to cause motor and non-motor deficits [2].

precise The pathological mechanism of dopaminergic neuronal degeneration is still several factors, includina unknown. but mitochondrial dysfunction, neuroinflammation, excitotoxicity, and abnormal protein aggregation, may contribute to the progression of PD [3]. The production of reactive oxygen species (ROS) reduces mitochondrial dysfunction by interfering with the electron transport chain (ETC), resulting in decreased ATP production and nigrostriatal dopaminergic neuronal loss [3].

Medicinal plants have been used as a natural source of bioactive compounds that provide therapeutic benefits and low-cost treatments for various diseases. Starchytapheta jamaicensis, a medicinal plant, has been reported to contain secondary metabolites with various medicinal properties, including analgesic, antidiarrheal, antimicrobial, antioxidant, antihypertensive, antinociceptive, and anti-inflammatory properties [4]. The plant contains several major groups of secondary metabolites, including alkaloids, flavonoids, phenols, steroids, and terpenoids. These bioactive compounds can be found in abundance throughout the plant. Álvarez et al. demonstrated that S. jamaicensis leaf ethyl acetate extract significantly suppresses reactive oxygen species (ROS) production by inhibiting XO and scavenging ROS [5]. Hexane extract, on the other hand, has no antioxidant activity.

Studies have extensively demonstrated that S. jamaicensis has antioxidant properties, which could be due to the presence of catechins (a type of flavonoid) in the leaves. Methanolic extract was found to be more effective in total antioxidant activity, DPPH, and FRAP than the other extracts [6]. An ethanol leaf extract of S. jamaicensis was also found to improve catalase activity in alloxan-induced diabetic rats [7].

The discovery of the first missense mutation in the SNCA gene, A53T, in 1997 [8], and the insoluble aggregated -Syn forms as the major component of LBs, a pathological hallmark of Parkinson's disease [9] has led to a new era in PD research [10]. Since then, more SNCA pathogenic mutations as well as multiplications of SNCA have been identified as genetic causes of PD.

Using the conventional Gal4/USA expression system, Feany and Bender created -Syn

transgenic Drosophila models by overexpressing either wild-type or familial mutants A53T and A30P of human -Syn [11]. These models replicate the key features of Parkinson's disease (PD): adult-onset loss of DA neurons, filamentous intra-neuronal inclusions containing -Syn, and locomotor dysfunction [12].

However, Parkinson's disease is a complex neurodegenerative disorder that requires further research into its underlying mechanisms and potential therapeutic targets. Starchytapheta jamaicensis has been shown to possess antioxidant properties and may have potential therapeutic benefits for the treatment of Parkinson's disease. Further studies are needed to investigate the efficacy and safety of S. jamaicensis as a potential treatment for Parkinson's disease.

In this study, we investigate the potential therapeutic benefits of S. jamaicensis leaf extracts in Parkinson's disease using Drosophila melanogaster as a model system. The Drosophila model is widely used to study the pathogenesis of Parkinson's disease due to its genetic tractability and conservation of key molecular mechanisms with humans [11,12]. Our study aims to explore the potential therapeutic benefits of S. jamaicensis leaf extracts in reducing oxidative stress and neurodegeneration Drosophila melanogaster in models of Parkinson's disease.

#### 2. MATERIALS AND METHODS

#### 2.1 Reagents

Analytical grade substances were used in this study. Acetylcholine iodide, DTNB, GSH, CDNB, sodium dihydrogen phosphate, potassium hydrogen phosphate, Randox protein kit, and EDTA were purchased from Sigma Aldrich and Chidex Surgical Suppliers Limited.

#### **2.2 Plant Collection and Extraction**

S. jamaicensis leaf was collected from the University of Calabar, Nigeria. The leaf was airdried for 7 days, then pounded and extracted in 90% ethanol for 72 hours. The extract was filtered and freeze-dried, and the percentage total yield was calculated using the formula: % extraction yield =  $(W2 - W1) / W0 \times 100$ .

#### 2.3 Drosophila Melanogaster Model

Transgenic fly lines expressing wild-type human synuclein (h- $\alpha$ S) under UAS control in neurons

were obtained from the Bloomington Drosophila Stock Center. When crossed with GAL4-elav.L females, the males expressed human alpha synuclein in neurons.

# 2.4 7-days LC50 and 28-days Survival Assay

The 7-day LC50 of the plant extract was tested using the ingestion method of exposure. Mortality readings were taken every 24 hours for 7 days. A survival study was performed using the same method, with mortality readings taken at 28 days.

### 2.5 Seven (7)-days Treatment of *D. melanogaster* with S. jamaicensis

Short exposure of D. melanogaster was conducted using the ingestion method of exposure. Treatment duration was determined from the survival study. Fifty flies were treated with low dose (LD) or high dose (HD) of the plant extract for 7 days. Mortality readings were taken every 24 hours.

### 2.6 Drosophila Culture and Treatment

Flies were cultured on standard Drosophila food containing agar, corn meal, sugar, and yeast. The PD flies were exposed separately to different doses of crude extract (Sigma Aldrich, CAS 458-37-7) and mixed in culture medium.

#### 2.7 Lifespan Determination

Newly enclosed male flies (control and PD) were placed in culture tubes containing 100, 200, and  $250\mu$ M of plant extract mixed in diet.

Flies were switched to a new diet every third day, and the number of dead flies was counted every three days until the last one died.

#### 2.8 Negative Geotaxis (measured by % of climbing activity)

Negative geotaxis was performed by applying the technique described by Abolaji et al. [13]. Ten flies from each group were counted every day for seven days.

#### 2.9 Acetylcholinesterase Activity

AChE activity was assessed using the Ellman method with minor adjustments. Protein content was employed to adjust the results as data were compared to the blank and sample blank.

#### 2.10 Estimation of Protein Carbonyl Content

The protein carbonyl content was estimated using the method described by Hawkins et al. [14]. The brain homogenate was diluted to a protein concentration of approx 1 mg/ml. About 250  $\mu$ L of each diluted homogenate was taken in an eppendorf centrifuge tube separately.

### 2.11 Total Thiol Determination

Total thiol determination was evaluated using the Ellman approach. The protocol used by Ololade et al. [15] was followed for the production of the assay reaction mixture and absorbance measurement.

### 2.12 Catalase Activity

Catalase activity was measured using Aebi's approach [16]. H2O2 clearance was monitored at 240 nm and 25°C while 10 L of the sample was mixed with solution A.

### 2.13 Malondialdehyde (MDA)

MDA was estimated using the method described by Hawkins et al. [14]. The pink-colored product's absorbance was measured at 532 nm after it had cooled.

# 2.14 Statistical Analysis

One-way analysis of variance (ANOVA) followed by Tukey's post-hoc test was used to find statistically significant differences between numerous groups receiving different treatments. Kaplan-Meier analysis method was performed to examine survival data, and log rank test was employed to compare groups. Statistical significance was defined as a P value of less than 0.05, 0.01 and 0.005 (p < 0.05, p < 0.01, p < 0.005).

#### 3. RESULTS AND DISCUSSION

# 3.1 Results

#### 3.1.1 Survival assay

When compared to the other groups, the number of dead flies in the transgenic group increased significantly and progressively over the course of the trial. Treatment with low and high doses of plant extract decreased transgenic flies' mortality and increased the number of flies that survived to the end of the experiment. This suggests the effect of S. jamaicensis leaf extract on toxicity in Transgenic Drosophila model of Parkinson's disease (PD) flies as seen in (Fig. 1).

#### 3.1.2 Negative geotaxis assay

The data from this result shows a significant decrease of locomotory activity in the transgenic (Sgg) group when compared to the wild treated (Wild+ plant extract) group, Low dose (LD) (Sgg+100mg/kg) and High dose (HD) (Sgg+250mg/kg) and no significance when compared to the Wild. (\*p<0.05, \*\*p<0.01, \*\*\*p<0.005). This suggests the Effect of S. iamaicensis leaf extract on Impaired motor function or mobility in Transgenic Drosophila model of Parkinson's disease (PD) flies as shown in (Fig. 2).

#### 3.1.3 Protein status

#### 3.1.3.1 Total protein level

The data from this result shows a significant increase of total protein level in the transgenic (Sgg+250mg/kg) group when compared to the wild treated (Wild+plant extract), transgenic (Sgg-) and Low dose (LD) (Sgg+100mg/kg) groups and no significance when compared to the Wild group. (\*\*p<0.01; \*\*\*p<0.005). This suggests the Effect of S. jamaicensis leaf extract on protein damage or degradation caused by oxidative stress in Transgenic Drosophila model of Parkinson's disease (PD) flies and seen in (Fig. 3).

#### 3.1.4 Lipid peroxidation

The data from this result shows a significant increase of malondialdehyde level in the transgenic (Sgg) group when compared to all other groups. (\*p<0.05, \*\*p<0.01, \*\*\*p<0.005). This suggests the effect of S. jamaicensis leaf extract on lipid peroxidation and oxidative damage to cell membranes in Transgenic Drosophila model of Parkinson's disease (PD) flies as seen in (Fig. 4).

#### 3.1.5 Oxidative stress parameters

#### 3.1.5.1 Total thiol level

The data from this result shows a significant increase of total thiol levels in the transgenic high dose (HD) (Sgg++250mg/kg) group when

compared to the Low dose (LD) (Sgg+100mg/kg) and a significance decrease when compared to the Wild. (\*p<0.05, \*\*p<0.01, \*\*\*p<0.005, \*\*\*\*p<0.001). This suggests the Effect of S. jamaicensis leaf extract on oxidative stress in Transgenic Drosophila model of 13428 Parkinson's disease (PD) flies as seen in (Fig. 5).

#### 3.1.5.2 Total catalase concentration

The data from this result shows a significant decrease of catalase concentration in the transgenic (Sgg+100mg/kg) group when compared to the wild, wild treated (Wild+ plant group and High dose (HD) extract) (Sgg++250mg/kg). (\*p<0.05, \*\*\*p<0.005). This suggests the Effect of S. jamaicensis leaf extract on alterations in cellular defenses against oxidative stress in Transgenic Drosophila model of Parkinson's disease (PD) flies as seen in (Fig. 6).

#### 3.1.6 Antioxidant enzyme activity

The data from this result shows a significant decrease of Acetylcholinesterase activity level in the transgenic (Sgg) group when compared to the *wild, wild treated (Wild+ plant extract),* Low dose (LD) (Sgg+50mg/kg) and High dose (LD) (Sgg+200mg/kg). (\*p<0.05, \*\*p<0.01, \*\*\*p<0.005). This suggests the Effect of S. jamaicensis leaf extract on acetylcholinesterase activity oxidative stress in Transgenic Drosophila

model of Parkinson's disease (PD) flies as seen in (Fig. 7).

#### 3.2 Discussion

The results of this study demonstrate that the S. jamaicensis extract exhibits a significant impact on the survival rate of D. melanogaster. The 7day LC50 value of 353 mg extract/10 g diet highlights the extract's toxicity, which is consistent with previous research that has identified various plant extracts as sources of toxic compounds affecting insect physiology [17]. Moreover, the significant reduction in the 28-day survival ratio of the treated flies compared to the control group (P < 0.05) underscores the detrimental effects of prolonged exposure to the extract. These findings suggest that the extract exerts a time-dependent negative effect on the viability of D. melanogaster, pointing to its potential toxicity.

In addition, we investigated the impact of the plant extract on AChE and climbing activities in D. melanogaster. AChE activity remains a prime marker of neurotoxicity, as its inhibition may indicate poor locomotive operations and common toxicity [18]. Our results showed that S. jamaicensis leaf extract did not significantly inhibit AChE and climbing activities when compared to control (P > 0.05). This suggests that the extract does not interfere with the antioxidant system of fruit flies [19].

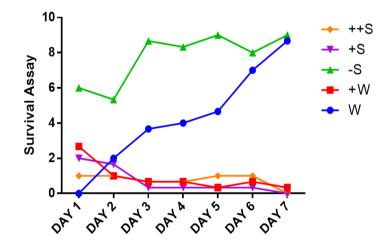


Fig. 1. Mortality rate of flies treated with different doses of S. jamaicensis leaf extract, per day for the duration of 7 days

Transgenic (Sgg) group: Significant from wild treated (Wild+ plant extract), Low dose (LD), and high dose group not Significant from wild group.

Wild (W), wild treated (+W), transgenic only group (-S), low dose (+S) high dose (++S)

Isamoh et al.; Asian J. Res. Rep. Neurol., vol. 7, no. 1, pp. 154-164, 2024; Article no.AJORRIN.124031

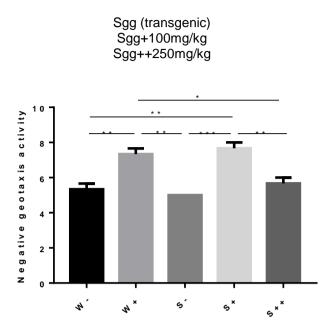
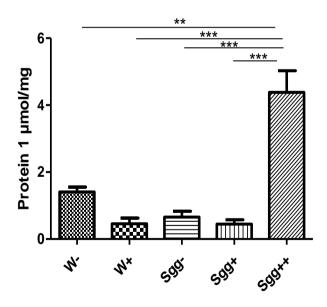


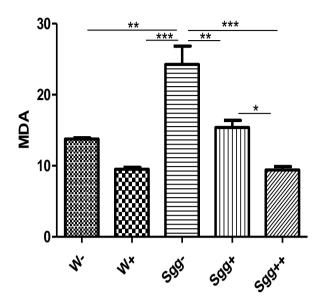
Fig. 2. Locomotory activity in transgenic Drosophila model of Parkinson's disease (PD) flies treated with different doses of S. jamaicensis leaf extract

Transgenic (Sgg) group: Significant from wild treated (Wild+ plant extract), Low dose (LD), and High dose group, and not Significant from Wild group. Data are presented as mean ± SEM (n=6). \*p<0.05; \*\*p<0.01; and \*\*\*p<0.005 compared the wild treated (Wild+ plant extract), transgenic (Sgg-), low dose and wild. Wild (W-), wild treated (W+), transgenic only group (Sgg-), low dose (Sgg+) high 13428dose (Sgg++)



# Fig. 3. Total protein levels in transgenic Drosophila model of Parkinson's disease (PD) flies treated with different doses of S. jamaicensis leaf extract

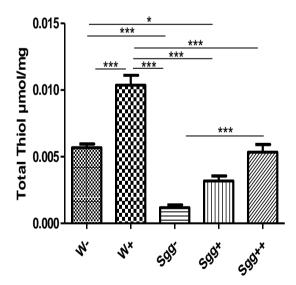
transgenic (Sgg+250mg/kg) group: Significant from wild treated (Wild+ plant extract) transgenic (Sgg-), Low dose (LD) group and not Significant from Wild group. Data are presented as mean ± SEM (n=6). \*p<0.05; \*\*p<0.01; and \*\*\*p<0.005 compared the wild treated (Wild+ plant extract), transgenic (Sgg-), low dose and wild. Wild (W-), wild treated (W+), transgenic only group (Sgg-), low dose (Sgg+) high dose (Sgg++). 134 Isamoh et al.; Asian J. Res. Rep. Neurol., vol. 7, no. 1, pp. 154-164, 2024; Article no.AJORRIN.124031



# Fig. 4. Malondialdehyde (MDA) levels in transgenic Drosophila model of Parkinson's disease (PD) flies treated with different doses of S. jamaicensis leaf extract

Transgenic (only) group: Significant from wild, wild treated (Wild+ plant extract) Low dose (LD) and High dose. Data are presented as mean  $\pm$  SEM (n=6). \*p<0.05; \*\*p<0.01; and \*\*\*p<0.005 compared the wild, wild treated (Wild+ plant extract), low dose and high dose groups.

Wild (W-), wild treated (W+), transgenic only group (Sgg-), low dose (Sgg+) high dose (Sgg++).



# Fig. 5. Total Thiol Level in transgenic Drosophila model of Parkinson's disease (PD) flies treated with different doses of S. jamaicensis leaf extract

Transgenic high dose (HD) (Sgg++250mg/kg) group: Significant from Low dose (LD) group. Data are presented as mean ± SEM (n=6). \*p<0.05; \*\*p<0.01; and \*\*\*p<0.005 compared with the wild, wild treated (Wild+ plant extract), and low dose groups.

Transgenic high dose (HD) (Sgg++250mg/kg) group: Significant from Wild group . 134Data are presented as mean ± SEM (n=6). \*p<0.05; \*\*p<0.01; and \*\*\*p<0.005 compared with the wild, wild treated (Wild+ plant extract), and low dose groups.

Wild (W-), wild treated (W+), transgenic only group (Sgg-), low dose (Sgg+) high dose (Sgg++).

Isamoh et al.; Asian J. Res. Rep. Neurol., vol. 7, no. 1, pp. 154-164, 2024; Article no.AJORRIN.124031

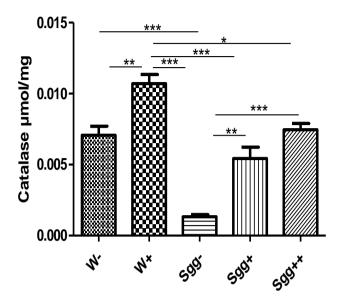


Fig. 6. Catalase concentration in transgenic Drosophila model of Parkinson's disease (PD) flies treated with different doses of S. jamaicensis leaf extract

transgenic (Sgg+100mg/kg) group: Significant from wild, wild treated (Wild+ plant extract), transgenic (only) group and High dose. Data are presented as mean ± SEM (n=6). \*p<0.05; \*\*p<0.01; and \*\*\*p<0.005 compared the wild, wild treated (Wild+ plant extract), and high dose groups.

Wild (W-), wild treated (W+), transgenic only group (Sgg-), low dose (Sgg+) high dose (Sgg++).

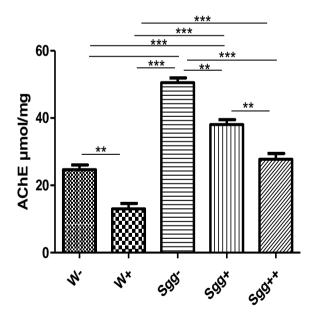


Fig. 7. Acetylcholinesterase (AChE) concentration in transgenic Drosophila model of Parkinson's disease (PD) flies treated with different doses of S. jamaicensis leaf extract (low dose 50 and high dose 200 mg/kg).

Transgenic (only) (Sgg-) group: Significant from wild, wild treated (Wild+ plant extract) Low dose (LD) and High dose. Data are presented as mean ± SEM (n=6). \*p<0.05; \*\*p<0.01; and \*\*\*p<0.005 compared to the wild, wild treated (Wild+ plant extract), low dose and high dose groups.

Wild (W-), wild treated (W+), transgenic only group (Sgg-), low dose (Sgg+) high dose (Sgg++).

Furthermore, our study demonstrated that the plant extract exhibits antioxidant activities in vivo, as shown by a significant increase in catalase (CAT) activities and whole thiol content compared to the control group. This suggests that the plant extract does not interfere with the antioxidant system of fruit flies, which is crucial for combating free radicals and maintaining cellular integrity [13].

The findings suggest that S. jamaicensis may have potential as a treatment for conditions related to oxidative stress-induced neuronal imbalance. The observed effects in fruit flies are particularly noteworthy due to the conservation of genes and biochemical pathways related to oxidative stress between fruit flies and humans [15]. This implies that the beneficial effects observed in fruit flies could potentially translate to humans.

Transgenic Drosophila model of Parkinson's disease may not fully capture the complex pathology and heterogeneity of human PD, which could impact the generalizability of our results. Furthermore, the efficacy of S. jamaicensis extract in reducing oxidative stress and neurodegeneration in PD may be specific to this Drosophila model and may not translate to humans.

We also did not investigate the specific mechanisms by which S. jamaicensis extract exerts its neuroprotective effects. Future studies must aim to elucidate the molecular targets and signaling pathways involved in its therapeutic effects, which is crucial for understanding its potential therapeutic benefits. Moreover, future research should also investigate the role of glutathione (GSH) as an antioxidant marker in the context of S. jamaicensis extract's neuroprotective effects. GSH is a critical antioxidant enzyme that plays a key role in protecting cells against oxidative stress [20]. In PD, GSH levels have been shown to be depleted [21], contributing to oxidative stress and neurodegeneration.

In addition, it would be essential to explore the effects of S. jamaicensis extract on dopaminergic neurons in the brain or midbrain using histology or immunohistochemistry of tyrosine hydroxylase (TH), a marker of dopaminergic neurons. This would provide valuable insights into the potential therapeutic benefits of S. jamaicensis extract in preserving dopaminergic neurons and preventing neurodegeneration in PD. Previous studies have shown that TH-positive neurons are significantly

reduced in the substantia nigra pars compacta of PD patients [22], making TH a valuable marker for investigating neurodegeneration in PD.

Furthermore, histological analysis of brain tissue provide valuable insights into the can morphology and distribution of dopaminergic neurons in response to S. jamaicensis extract treatment. This could be achieved using techniques such as immunohistochemistry or fluorescent staining for TH or other dopamine-Additionally, related markers [23]. immunohistochemical analysis of brain tissue can also help identify potential changes in neuronal morphology, such as increased oxidative stress or inflammation, that may be indicative of neurodegenerative changes [24].

While this study has limitations, it provides valuable insights into the potential therapeutic benefits of S. jamaicensis extract in PD. Future studies should strive to address these limitations and explore the translational potential of this natural compound, ultimately bringing us closer to its potential clinical application. Specifically, further research should aim to elucidate the molecular mechanisms underlying S. jamaicensis extract's neuroprotective effects, including the role of GSH as an antioxidant marker, and investigate its effects on dopaminergic neurons in the brain or midbrain using histology or immunohistochemistry of TH.

# 4. CONCLUSION

In conclusion, our study demonstrates that the ethanol extract of S. jamaicensis leaf shows a trend towards increasing total thiol and catalase levels in D. melanogaster, although these changes were not statistically significant (P > 0.05). Despite this, the extract significantly enhances the survival rate, young fly emergence rate, and climbing activities (acetylcholinesterase and negative geotaxis) of adult D. melanogaster treated with the extract.

Overall, these findings suggest that S. jamaicensis may have potential as a treatment for conditions related to oxidative stress-induced neuronal imbalance. Further research is needed to elucidate the specific mechanisms involved and to determine the potential therapeutic implications of these findings.

#### DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative Al technologies such as Large Language Models

(ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

### CONSENT

It is not applicable.

#### ETHICAL APPROVAL

This study was conducted in accordance with the guidelines of the College of medical sciences ethical committee, university of calabar, which approved the use of Drosophila has melanogaster in research. The protocols were reviewed and approved and the ethical approval number was obtained 202ANA1023. The flies were handled and maintained in accordance with the guidelines for animal care and welfare, and all efforts were made to minimize their stress and discomfort.

An approval for the experimental protocol of this study with registration number 190ANA3023 was obtained from the Faculty Animal Research Ethics committee, Faculty of Basic Medical Sciences, University of Calabar, Cross River state, Nigeria.

# **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

# REFERENCES

 Ramesh S, Arachchige ASPM. Depletion of dopamine in Parkinson's disease and relevant therapeutic options: A review of the literature. AIMS Neurosci. 2023;10(3): 200-231.

DOI: 10.3934/Neuroscience.2023017.

- Magrinelli F, Picelli A, Tocco P, Federico A, Roncari L, Smania N, Zanette G, Tamburin S. Pathophysiology of Motor Dysfunction in Parkinson's Disease as the Rationale for Drug Treatment and Rehabilitation. Parkinsons Dis. 2016;2016:9832839. DOI: 10.1155/2016/9832839.
- Dexter DT, Jenner P. Parkinson disease: from pathology to molecular disease mechanisms. Free Radic Biol Med. 2013; 62:132–144.
- Idu M, Omogbai EKI, Aghimien GE, Amaechina F, Timothy O, Omonigho SE. Preliminary phytochemistry, antimicrobial properties and acute toxicity of Stachytarpheta jamaicensis (L.) Vahl.

leaves. Trends in Medical Research. 2007; 2(4):193–198.

DOI: 10.3923/tmr.2007.193.198.

- Álvarez E, Leiro JM, Rodríguez M, Orallo F. Inhibitory effects of leaf extracts of Stachytarpheta jamaicensis (Verbenaceae) on the respiratory burst of rat macrophages. Phytotherapy Research. 2004;18(6):457–462. DOI: 10.1002/ptr.1442.
- Liew PM, Yong YK. Stachytarpheta jamaicensis (L.) Vahl: From Traditional Usage to Pharmacological Evidence. Evid Based Complement Alternat Med. 2016; 2016:7842340. DOI: 10.1155/2016/7842340.
- Wan Rozianoor MH, Nurol Eizzatie Y, Nurdiana S. Hypoglycemic and antioxidant activities of Stachytarpheta jamaicensis ethanolic leaves extract on alloxan-induced diabetic sprague dawley rats. BioTechnology. 2014;9(10):423–428.
- Polymeropoulos MH, Lavedan C, Leroy E, Ide SE, Dehejia A, Dutra A, Pike B, Root H, Rubenstein J, Boyer R, Stenroos ES, Chandrasekharappa S, Athanassiadou A, Papapetropoulos T, Johnson WG, Lazzarini AM, Duvoisin RC, Di Iorio G, Golbe LI, Nussbaum RL. Mutation in the alpha-synuclein gene identified in families with Parkinson's disease. Science. 1997; 276(5321):2045-7.

DOI: 10.1126/science.276.5321.2045.

- Spillantini MG, Schmidt ML, Lee VM, Trojanowski JQ, Jakes R, Goedert M. Alpha-synuclein in Lewy bodies. Nature. 1997;388(6645):839-40. DOI: 10.1038/42166.
- Sian-Hulsmann J, Riederer P. The Nigral Coup in Parkinson's Disease by α-Synuclein and Its Associated Rebels. Cells. 2021;10(3):598. DOI: 10.3390/cells10030598.
- Feany MB, Bender WW. A Drosophila model of Parkinson's disease. Nature. 2000;404:394–8. DOI: 10.1038/35006074.
- 12. Rahul and Siddique YH. Drosophila: a model to study the pathogenesis of Parkinson's disease. CNS Neurol. Disord. Drug Targets. 2022;21:259–277.
- Shulman JM, De Jager PL, Feany MB. Parkinson's disease: genetics and pathogenesis. Annu Rev Pathol. 2011; 6:193-222. DOI:10.1146/annurev-pathol-011110-130242.

- Abolaji AO, Kamdem JP, Lugokenski TH, Nascimento TK, Waczuk EP, Farombi EO. Involvement of oxidative stress in 4vinylcyclohexene-induced toxicity in Drosophila melanogaster. Free Radic Biol Med. 2019;71:99–108.
- 15. Hawkins CL, Morgan PE, Davies MJ. Quantification of protein modification by oxidants. Free Radic Biol Med. 2009; 46:965–988.
- Ololade, Zacchaeus, Ololade, Z.S. and Olawore, N.O. Characterization of Essential Oil from the Seed of Eucalyptus cloeziana and Evaluation of its Modes of Medicinal Potentials, Edorium Journal of Infectious Diseases. 2017; 3, 1-8.
- Aebi H. Catalase in vitro. Methods Enzymol. 1984;105:121-6. DOI:10.1016/s0076-6879(84)05016-3. PMID: 6727660.
- 18. Alexander EM, Aguiyi JC, Mdekera IW, Ogwu OS, Imoleavo OO, Ugokwe CV. The Climbina Performance, Neuromuscular Transmitter (ACHE) Activity, Reproductive Performance and Survival of Drosophila melanogaster Fed Diet with Mangiferaindica Cold Aqueous Leaf Extract. J Appl Life Sci Int. 2019;22(2):1-11.
- Kavitha P, Rao JV. Oxidative stress and locomotor behaviour response as biomarkers for assessing recovery status of mosquito fish, Gambusia affinis after lethal effect of an organophosphate pesticide, monocrotophos. Pestic Biochem Physiol. 2007;87(2):182–8.

20. Vašková J, Kočan L, Vaško L, Perjési P. Glutathione-Related Enzymes and Proteins: A Review. Molecules. 2023;28(3): 1447.

DOI: 10.3390/molecules28031447.

- Leathem A, Ortiz-Cerda T, Dennis JM, Witting PK. Evidence for Oxidative Pathways in the Pathogenesis of PD: Are Antioxidants Candidate Drugs to Ameliorate Disease Progression? Int J Mol Sci. 2022;23(13):6923. DOI: 10.3390/ijms23136923.
- Roostalu U, Salinas CBG, Thorbek DD, Skytte JL, Fabricius K, Barkholt P, John LM, Jurtz VI, Knudsen LB, Jelsing J, Vrang N, Hansen HH, Hecksher-Sørensen J. Quantitative whole-brain 3D imaging of tyrosine hydroxylase-labeled neuron architecture in the mouse MPTP model of Parkinson's disease. Dis Model Mech. 2019;12(11):dmm042200. DOI: 10.1242/dmm.042200.
- Kim D, Bak MS, Park H, Baek IS, Chung G, Park JH, Ahn S, Park SY, Bae H, Park HJ, Kim SK. An Automated Cell Detection Method for TH-positive Dopaminergic Neurons in a Mouse Model of Parkinson's Disease Using Convolutional Neural Networks. Exp Neurobiol. 2023;32(3):181-194.

DOI: 10.5607/en23001.

24. Sidorova Y, Domanskyi A. Detecting Oxidative Stress Biomarkers in Neurodegenerative Disease Models and Patients. Methods Protoc. 2020;3(4):66. DOI: 10.3390/mps3040066.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of the publisher and/or the editor(s). This publisher and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/124031